

REVIEW

Open Access



# Natural and socio-environmental factors in the transmission of COVID-19: a comprehensive analysis of epidemiology and mechanisms

Zhaoyuan Gong<sup>1†</sup>, Tian Song<sup>1†</sup>, Mingzhi Hu<sup>1†</sup>, Qianzi Che<sup>1</sup>, Jing Guo<sup>1</sup>, Haili Zhang<sup>1</sup>, Huizhen Li<sup>1</sup>, Yanping Wang<sup>1\*</sup>, Bin Liu<sup>1\*</sup> and Nannan Shi<sup>1\*</sup>

## Abstract

**Purpose of review** There are significant differences in the transmission rate and mortality rate of COVID-19 under environmental conditions such as seasons and climates. However, the impact of environmental factors on the role of the COVID-19 pandemic and the transmission mechanism of the SARS-CoV-2 is unclear. Therefore, a comprehensive understanding of the impact of environmental factors on COVID-19 can provide innovative insights for global epidemic prevention and control policies and COVID-19 related research. This review summarizes the evidence of the impact of different natural and social environmental factors on the transmission of COVID-19 through a comprehensive analysis of epidemiology and mechanism research. This will provide innovative inspiration for global epidemic prevention and control policies and provide reference for similar infectious diseases that may emerge in the future.

**Recent findings** Evidence reveals mechanisms by which natural environmental factors influence the transmission of COVID-19, including (i) virus survival and transport, (ii) immune system damage, (iii) inflammation, oxidative stress, and cell death, and (iiii) increasing risk of complications. All of these measures appear to be effective in controlling the spread or mortality of COVID-19: (1) reducing air pollution levels, (2) rational use of ozone disinfection and medical ozone therapy, (3) rational exposure to sunlight, (4) scientific ventilation and maintenance of indoor temperature and humidity, (5) control of population density, and (6) control of population movement. Our review indicates that with the continuous mutation of SARS-CoV-2, high temperature, high humidity, low air pollution levels, and low population density more likely to slow down the spread of the virus.

**Keywords** COVID-19, Nature environmental factor, Social environmental factor, Epidemiology, Mechanism research

<sup>†</sup>Zhaoyuan Gong, Tian Song and Mingzhi Hu contributed equally to this work.

\*Correspondence:

Yanping Wang  
wangyanping4816@163.com

Bin Liu  
lynch1123@126.com

Nannan Shi  
13811839164@vip.126.com

Full list of author information is available at the end of the article



## Introduction

Coronavirus disease 2019 (COVID-19) is defined as a disease caused by the Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2), which is an emerging respiratory infection. COVID-19 can occur through close contact with an infected person. The disease is characterized by high transmission rate, long incubation period, and global spread. On 11 March 2020, this disease was declared a global pandemic by the World Health Organization (WHO) [1]. The outbreak of COVID-19 has caused many threats and dangers to human health, including increased mortality and morbidity globally [2, 3]. The COVID-19 pandemic is challenging the world economy and health systems and demonstrates the extent of global interdependence and the need to address global health threats [4].

Current research on COVID-19 mainly focuses on vaccines, viruses, hosts and drugs [5]. In contrast, research on the impact of environmental factors on COVID-19 needs to be further carried out. The incidence of many similar infectious diseases showed seasonal patterns, including human coronaviruses [6]. COVID-19 is no exception and may have seasonal epidemic peaks. Therefore, understanding which environmental factors influence COVID-19 can allow planning and implementing public health interventions and capacities to reduce the impact of the disease. It will provide innovative inspiration for global epidemic prevention and control policies and provide reference for similar infectious diseases that may emerge in the future.

Although SARS-CoV-2 transmission has been recorded in almost all countries, there is significant spatial and temporal heterogeneity in transmission dynamics, morbidity and mortality across countries, regions and even communities [7]. Many studies have investigated the correlation between this spatial and temporal heterogeneity and environmental factors, including natural environmental factors and social environmental factors [8]. Direct or indirect evidence shows that these environmental factors have an impact on the spread and development of COVID-19 [9].

This research is used in Scopus, ISI scientific network and PubMed database ("climate" OR "climate change" OR "temperature" OR "precipitation" OR "relative humidity" OR "wind speed" OR "sunlight" OR "wind speed" OR "water resources" OR "solar radiation" OR "social environment factor" OR "air pollution" OR "PM2.5" OR "PM10" OR "ozone" OR "NO<sub>2</sub>" OR "CO" OR "wastewater" OR "heavy metal pollution" OR "sociodemographic characteristics" OR "local policies" OR "socioeconomic activity") and ("COVID" OR "Coronavirus disease 2019" OR "COVID-19" OR "SARS-CoV-2" OR "Novel Coronavirus" OR "COVID-19 transmission" OR " Novel

Coronavirus transmission" OR "COVID-19 confirm case"). Inclusion criteria are all relevant manuscripts that assess the impact of environmental factors on the number of cases and incidence rate of COVID-19. Exclusion criteria include comments, letters, editorials, conference abstracts, and low-quality research.

This review summarizes the impact of environmental factors on the transmission of COVID-19 and summarizes studies from epidemiological evidence to mechanisms. We divide environmental factors into natural environmental factors and social environmental factors. Natural environmental factors include air pollution, temperature, humidity, wind speed, rainfall, solar radiation, soil and water resources. Social environmental factors include sociodemographic characteristics, local policies and socioeconomic activity. Finally, there is a discussion of the potential future directions in this field. Critical assessments of these relationships can enhance estimates of the risk of similar infectious diseases from environmental exposures and guide the design of interventions to slow the spread of the virus and protect vulnerable populations from infection. This will provide innovative inspiration for global epidemic prevention and control policies and provide reference for similar infectious diseases that may emerge in the future.

## Natural environment factors and transmission of COVID-19

### Native environment factor

#### *Temperature and humidity*

The relationship between viral infections and meteorological conditions has been of concern in the past. Researchers from China looked at the link between temperature and humidity in more than 3750,000 confirmed COVID-19 cases from 185 countries. It found that 60.0% of confirmed COVID-19 cases occurred in places with temperatures between 5°C and 15°C, with a peak of 11.54°C. In addition, about 73.8% of confirmed cases are concentrated in areas with absolute humidity of 3 g/m<sup>3</sup> to 10 g/m<sup>3</sup> [10]. In Japan, researchers conducted a longitudinal cohort study of 6,529 confirmed COVID-19 cases across 28 geographical areas. The results showed that the increase of the COVID-19 epidemic was significantly correlated with the increase of daily temperature or sunshine duration [11]. Haque and Rahman found that high temperature and high humidity significantly reduced the spread of COVID-19, respectively. In Bangladesh, more than four-fifths (84.2%) of the total cases were clustered within the average temperature range (26–28°C) [12]. This is consistent with the results of a previous ecological study, which found that the optimal ambient temperature associated with SARS cases was between 16°C and 28°C, based on data from Hong Kong, Guangzhou, Beijing and

Taiyuan [13]. The seasonal cycle of respiratory viral diseases has been widely recognized for thousands of years. The temporal trends of COVID-19 transmission presented a periodic fluctuation and reflected the seasonal changes. In terms of time series, in line with the seasons in Brazil, the average temperature and relative humidity from March to May (autumn) had a significant positive effect on new cases, while the months from June to August (winter) and September to November (spring) had a negative effect [14].

In a systematic review of 17 studies has found that climate parameters may be an important factor in the spread of COVID-19. Cold and dry conditions enhance the spread of the virus [15]. A systematic review of 62 publications were published between December 2019 and February 2021 on the association between climate factors and the spread of COVID-19, found consistent results that high temperatures may have significantly influenced the spread of COVID-19 and suppressed the pandemic [16]. Yuan et al. found that they may be negatively associated with daily new cases of COVID-19 in 127 countries when temperature, relative humidity, and wind speed are lower than 20 °C, 70%, and 7 m/s, respectively. In a follow-up study, it found to be inversely associated with daily new cases of COVID-19 in 188 countries when temperatures and relative humidity below 21 °C and 64%, respectively. And in these two studies, the researchers found that temperature and humidity were negatively correlated with the daily number of new COVID-19 cases and deaths [17, 18]. In India, researchers found that most COVID-19 cases had surface temperatures between 24 and 30°C and relative humidity between 50 and 80 percent, which is highly dependent on relative humidity at certain temperatures [19]. However, some studies [20] report that SARS-CoV-2 transmission is ineffective with increasing temperature. For the role of meteorological parameters, this contradictory discovery is mainly due to the difference in analysis methods and limitation in data availability of each study.

#### ***Solar radiation, sunlight exposure, and wind speed***

Other climatological factors can also affect the spread of SARS-CoV-2, previous evidence shows that solar radiation and wind speed also affect the spread of infectious diseases [21, 22]. For example, in the tropical state of Rio de Janeiro in Brazil, studies have found that high solar radiation can be shown to be a major climatic factor in curbing the spread of COVID-19. Solar radiation was positively correlated with the infection of COVID-19. There was a significant negative correlation between higher wind speed and lower incidence of COVID-19 [23]. Researchers in Iran found that areas with low wind speeds and exposure to solar radiation had higher

rates of infection, which helped the virus survive [24]. In a descriptive observational cross-sectional study conducted in France, a significant negative association between sun exposure and COVID-19 mortality was observed [25]. Another study analyzed the correlation between the meteorological parameters and the transmission of COVID-19 in Baghdad, the capital of Iraq. The results show that temperature, wind speed and solar radiation are the primary meteorological parameters leading to the spread of COVID-19 in Baghdad and are related to the confirmed cases and deaths of COVID-19 [26]. But a study on the correlation between solar radiation exposure and the COVID-19 pandemic in Jakarta, Indonesia, found different results. The study found no significant correlation between sun exposure and morbidity and death in patients with COVID-19. Sunlight was significantly associated with recovery from COVID-19 [27]. In the latest study, Al-Khateeb et al. compared the association between multiple regional weather conditions in the Northern hemispheres, Southern hemispheres, and Irbid, Jordan and COVID-19 transmission, found that the relationship between wind speed and spread of COVID-19 was oscillatory and insignificant on a worldwide [28]. Statistical results may not confirm a specific causal relationship between exposure to solar UV radiation and disease variables such as morbidity and mortality [29].

#### ***Water resource***

SARS-CoV-2 deposited on the surface of objects can be washed into the surface runoff by rainfall. Although the waterborne transmission of COVID-19 has not been confirmed, the potential risk cannot be ignored. Research therefore needs to answer whether hydrological conditions (such as river length, lake area, precipitation and volume of water resources) are related to COVID-19 outbreaks. For example, the researchers investigated the associations between hydrological factors such as lake area, river length, precipitation and volume of water resources in 30 regions of China and the incidence of COVID-19. The results showed that the number of confirmed COVID-19 cases was moderately correlated with river length and precipitation, but weakly correlated with water resources [30]. Precipitation had been analyzed in studies in Indonesia, the United States and Brazil, had not been found to be associated with COVID-19 [31–33]. However, other studies had also found that precipitation is positively correlated with the spread of COVID-19. Countries with higher rainfall showed an increase in disease transmission. On average, there were 56.01 additional cases per inch/day [33].

## Environmental pollution factor and transmission of COVID-19

### Air pollution

Epidemiologic evidence shows a strong link between air pollution and COVID-19, contributing significantly to the transmission and severity of COVID-19 [34, 35]. In the United States, researchers used ecological regression analysis to examine the relationship between long-term (2000–2016) average PM<sub>2.5</sub> concentrations and COVID-19 mortality over 3,089 counties. After accounting for many county-level confounders, researchers found that a 1 µg per m<sup>3</sup> increase in PM<sub>2.5</sub> was associated with an 11% increase in county-level COVID-19 mortality rate [36]. The same results were found in another study, in which researchers found a significant positive correlation between 2016 average PM<sub>2.5</sub> concentrations in 3,110 US counties and COVID-19 mortality [37]. An ecological study in Italy found a positive correlation between PM<sub>2.5</sub> concentrations and excess mortality associated with COVID-19 in Northern Italy. A one-unit increase in PM<sub>2.5</sub> concentration (µg/m<sup>3</sup>) is associated with a 9% increase in COVID-19 related mortality [38]. Several ecological studies in other countries such as China, UK and Netherlands had found the same results, with areas with poorer air quality are more likely to have elevated COVID-19 incidence and mortality [39–41].

Short-term exposure to air pollution may also affect COVID-19, with multiple studies finding that air pollution may affect recovery time, mortality, morbidity, and emergency department visits [42]. For example, researchers conducted a case-crossover study of 78,255 emergency department visits for COVID-19 in two Canadian provinces. The study found a significant correlation between PM<sub>2.5</sub> and emergency department visits for COVID-19 [43]. In Changsha, China, Liu et al. found that long recovery duration among COVID-19 patients was positively correlated with short-term exposure to PM<sub>2.5</sub>, NO<sub>2</sub>, and CO [44]. The researchers also found that COVID-19 patients with both Delta and Omicron had an increased chance of developing early respiratory COVID-19 manifestations after short-term exposure to air pollution [45].

In addition to particulate matter (PM), which has been widely studied, a number of studies have also involved carbon monoxide (CO), nitrogen dioxide (NO<sub>2</sub>), ozone (O<sub>3</sub>) and other air pollutants [46]. In a comparative study of the impact of air pollution on COVID-19 in multiple countries, a correlation between pollutant gases and COVID-19 risk was found in the United States, Italy and Spain, while in China the relationship was negative. Factors in air pollution have different associations with COVID-19 risk [47]. Another study looked at 446,440 COVID-19 cases, covering 4,609 census tracts in

southern California. The pooled RR (95%CI) for the incidence of COVID-19 associated with 1-year exposure to NO<sub>2</sub> and O<sub>3</sub> were 1.09 (1.02, 1.17) per 3.2 ppb and 1.06 (1.00, 1.12) per 5.5 ppb respectively [48]. In Los Angeles, long-term exposure to NO<sub>2</sub> has been associated with an increased risk of COVID-19 cases and mortality. The researchers found that an 8.7 ppb increase in annual mean NO<sub>2</sub> concentrations was associated with a 16–31% increase in the rate of COVID-19 cases and a 35–60% increase in mortality [49].

### Wastewater

The waterborne transmission of COVID-19 has not been confirmed. Early in the SARS-CoV-2 outbreak, it was reported that live SARS-CoV-2 could be isolated from the feces and urine of COVID-19 patients [50]. And according to the experience accumulated during the SARS epidemic, sewage systems could also be contaminated by the virus [51]. Different concentrations of SARS-CoV-2 have been detected in wastewater in the Netherlands, Belgium, Australia, and the United States [52–54]. To date, there have been no studies in the public domain on the persistence and survivability of SARS-CoV-2 in water or wastewater. But a link between the virus and persistence and survival can be found from previous studies of coronaviruses. For example, human coronavirus 229E can survive for 7 days in water at 23 °C [55]. Researchers performed SARS-CoV-2 RNA detection by real-time RT-PCR and infectivity test on culture cells on three river samples in Milan, Italy. Real-time RT-PCR results showed positive, but infectivity was not effective [56].

Through the secondary utilization of wastewater and the disposal of medical waste, SARS-CoV-2 can enter the soil environment [57, 58]. In China, 20% of soil samples taken near hospitals receiving COVID-19 subjects and Wuhan sewage treatment plants recently tested positive for SARS-CoV-2 RNA, with abundance ranging from 205 to 550 copies/g [59]. Similar to the research status of viruses in water resources, the activity and infectivity of viruses in soil resources have not been widely studied and confirmed [60]. But there is no denying that the virus can seriously affect soil health, and improper handling will pose a threat to human and animal health.

### Heavy metal pollution

Hydrosphere and pedosphere are essential natural environment factors. Since heavy metals are not biodegradable, heavy metal ions in water and in soil be biologically accumulated via the food chain towards the human body. Most heavy metals, such as arsenic (As), lead (Pb), mercury (Hg), and cadmium (Cd) are considered environmental pollutants [61]. Previous evidence suggested that heavy metal exposure is associated with



higher mortality from influenza or pneumonia [62]. In the laboratory, heavy metal exposure has been found to play a role in impaired mucociliary clearance, reduced barrier function, airway inflammation, oxidative stress, and apoptosis [63]. Exposure to these heavy metals after COVID-19 infection may increase the risk of severe COVID-19 through these abnormal or exaggerated immune responses [64]. Studies had found that exaggerated immune responses are associated with multiple organ system failure, COVID-19 hospitalization, and death [65].

Chronic exposure to As, Cd, Hg, and Pb has been associated with respiratory dysfunction and respiratory diseases [66]. Solenkova et al. reviewed English-speaking medical literature to find that Hg, Pb, Cd, and as are associated with cardiovascular disease of atherosclerotic origin [67]. In addition, epidemiologic studies have found that cumulative exposure to heavy metal mixtures is associated with obesity and its associated chronic diseases, such as hypertension and type 2 diabetes [68]. These diseases have a significant impact on COVID-19. For example, the most common comorbidities found in COVID-19 cases in clinical studies are hypertension, followed by diabetes. More comorbidities were associated with poorer clinical outcomes. Obesity and type 2 diabetes are risk factors for poor COVID-19 prognosis [69].

While it is true that heavy metals have impact on COVID-19 patients, there is a lack of direct data linking exposure to heavy metals to the risk and/or severity of COVID-19. In a retrospective analysis of 306 patients confirmed COVID-19 in China, researchers analyzed levels of essential and/or toxic metals (classes) in whole blood, depending on the severity and outcome of the disease. The results found that among severely ill patients, the death group had higher levels of chromium and cadmium and lower levels of arsenic compared to the recovery group [70]. One study found that COVID-19 patients with elevated levels of chromium, cadmium, mercury and lead in their urine had a poorer prognosis (severe and non-severe) [71].

### **Social environment factor and transmission of COVID-19** ***Evidence linking sociodemographic characteristics to transmission of COVID-19***

Droplet or airborne transmission is the main route of SARS-CoV-2 transmission, and higher population density often leads to the long-term spread of COVID-19 [72]. In Malaysia, areas with a high number of residents and high population density have a greater number of cases in proportion to the population of the area. The correlation between COVID-19 cases and population density was strongest in the central region [73]. Another study using long-term data on the relationship between

external demographic parameters such as total population, population density and weighted population density and the spread of COVID-19 in Malaysia found different results. The results showed that there was a strong and significant positive correlation between total population and COVID-19 cases. However, a weak positive relationship was found between density variables (population density and weighted population density) and the spread of COVID-19 [74]. But most studies show that increasing population density in turn leads to an increase in COVID-19 cases and deaths [75]. Population density is thought to have a more significant impact on COVID-19 than meteorological factors. For example, the researchers investigated the correlation of spread and decay durations of the COVID-19 pandemic in China, the United Kingdom, Germany, and Japan with temperature, humidity, and population density. The results showed that propagation duration and decay durations were significantly correlated with population density, and the effect of population density was more significant than that of meteorological factors [76]. The characteristics of the built environment at different spatial scales caused by different population parameters will also affect the prevention and spread of infectious diseases. Poor housing conditions and high building density can lead to problems with inadequate sanitation facilities, which will create an environment conducive to disease transmission [77]. In Hong Kong, China, for example, research had shown that high transport accessibility, dense high-rise buildings, higher density of commercial land, and a higher land use mix are associated with a higher risk of being visited by confirmed cases. More green space, higher median household income, and lower commercial land density were associated with a higher risk of housing with confirmed cases [78].

There are race-related health disparities in the COVID-19 pandemic, with higher morbidity and mortality rates among ethnic minorities. Black workers most affected by the outbreak are more likely to be employed in key industries, in occupations that involve frequent exposure to infections and close relationships with others [79]. But other studies had found little evidence that occupation affects infection rates. For example, infection rates among frontline healthcare workers have not been shown to be higher than those non frontline healthcare workers. The strongest risk factors associated with COVID-19 infection among health care workers were neighborhood infection rates and ethnicity [80]. In addition, risk factors for developing COVID-19 in adults include age and gender [81]. For example, Increased mortality from COVID-19 was significantly associated with higher rates of obesity in women and higher rates of smoking in men [82]. There is growing evidence that COVID-19 produces

more severe symptoms and higher mortality in men than in women [83]. As a vulnerable group with reduced immune system effectiveness, the elderly are often at a higher risk of infectious diseases [84]. Susceptibility to SARS-CoV-2 infection increases with age [85, 86].

#### ***Evidence linking local policies and socioeconomic activity to transmission of COVID-19***

Person-to-person transmission is the main way of transmission of COVID-19. In response to the threat of the epidemic, many countries have introduced measures to restrict the movement of people. In Wuhan, China, for example, there was a significant decrease in new cases during the four-day lockdown. During that time, the increase in new cases dropped by about 50%, with the number of cases fluctuating on the fifth day and then rapidly decreasing [87]. The researchers investigated the movement of people and government restrictions as a function during successive waves of SARS-CoV-2 mutation in Canada. The results showed that in the first two years, government restrictions were high, and turnover was low, characterizing a 'seek-and-destroy' approach. After this phase, the highly transmissible Omicron (B.1.1.529) variant began circulating in NS at the end of the following year, leading to an increase in cases, hospitalizations, and deaths. During Omicron, although the transmissibility (26.41 times) and lethality (9.62 times) of the new variant increased, unsustainable government restrictions and declining public compliance led to increased population mobility [88]. In another cross-sectional study, containment and confinement were found to be significantly associated with overall mobility and were associated with a reduction in SARS-CoV-2 infection [89]. Lockdowns also reduce air pollution, and NO<sub>2</sub> concentrations can be used as environmental indicators to evaluate the effectiveness of lockdowns. In the United Kingdom, researchers found that exposure to NO<sub>2</sub> dropped significantly during lockdowns, while exposure to PM<sub>2.5</sub> dropped relatively little [90]. In a Cochrane systematic review, 84 studies were analyzed and found that isolation or microbiological testing, or a combination of both, prevented further cases. These interventions may have a positive shift in the development of the epidemic, and case detection may improve [91]. It has also been reported that 100% use of masks combined with lockdown is a measure that can reduce the risk of additional waves [92]. Masks could be one of the main pillars in the fight against the virus [93]. Finally, with lockdown measures in place, maintaining adequate indoor air quality levels is critical to slowing airborne viruses [94].

Socioeconomic activity has also been the focus of many studies investigating the factors affecting COVID-19 [95]. Epidemics may accelerate during periods of economic

activity, possibly because of an increase in the number of people traveling, followed by an increase in human contact. For example, studies had proved that international trade exceeded other common parameters used to prove the spread of COVID-19 due to economic, demographic, environmental and climatic factors [96]. The sum of international data on import and export trade can be a complex but appropriate indicator for measuring the underlying socioeconomic dynamics of geo-economic areas [96]. Another study examined the role of trade in the dynamics of epidemic spread within and between countries in three large European countries: Italy, France, and Spain. The findings suggest that the association between trade and outbreak severity appears to be supported by empirical evidence, potentially introducing new hypotheses to explain the dynamics of COVID-19 transmission within and between countries [97].

#### **Mechanism of nature environmental factors influencing the SARS-CoV-2**

##### **Native environment factor**

##### ***Temperature and humidity***

As noted above, there is growing epidemiologic evidence that the risk of transmission of SARS-CoV-2 was influenced by temperature and humidity. Many studies at the molecular level may further confirm this idea. Temperature can promote changes in the molecular structure of biomacromolecules (i.e., nucleic acids, proteins, lipids) until affecting their function. In the case of proteins, temperature is known to induce changes in secondary and tertiary structures, resulting in structural alterations that alter their stability and their role in regulating cellular processes, signal transduction, and intrinsic enzyme properties [98]. For example, the researchers used molecular dynamics simulation (MD) to reveal the molecular basis of the effect of temperature on the SARS-CoV-2 spike glycoprotein. The results showed that temperature induced conformational change of S1 subunit of SARS-CoV-2 spike glycoprotein that remodel the internal hydrogen bonding structure and especially affected secondary structure of the main region of interaction (RBD) of the spike glycoprotein of SARS-CoV-2 with the human ACE2 receptor [99]. Relative humidity (RH) can be considered an extrinsic factor for viral stability, as it controls evaporation, which affects the size of viral droplets, their physical fate, and their chemical microenvironment [100]. Table 1 summarizes some key studies of the effects of temperature and humidity on SARS-CoV-2 survival on different substrates. We found that the virus can survive on many substrates, and the lower the temperature and humidity, the longer the half-life of the virus. In one study, the half-life of the SARS-CoV-2 virus at 4 °C was three times that at 22 °C. One of the studies in the

**Table 1** Study of temperature and humidity on the activity of SARS-CoV-2

Substrate	Virus	Virus titer	Type	Temperature (°C) and relative humidity (%)	Virus stability (hour (h))	Reference
Human-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	25 °C and 70%	t1/2: 2.3–12.57 h	[104]
Human-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	21 °C and 60%	t1/2: 5.23–16.74 h	[104]
Human-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	13 °C and 66%	t1/2: 15.98–54.34 h	[104]
Human-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	5 °C and 75%	t1/2: 33.37–121.83 h	[104]
Surfaces under indoor	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	25 °C and 70%	t1/2: 2.54–5.58h	[105]
Surfaces under indoor	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	21 °C and 60%	t1/2: 3.5–12.86h	[105]
Surfaces under indoor	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	13 °C and 66%	t1/2: 17.11–31.82h	[105]
Surfaces under indoor	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA1/2020	5 °C and 75%	t1/2: 47.94–121.78 h	[105]
Steel	Delta B.1.617.2	$1.1 \times 10^4$ pfu	HCM/V/078	Room temperature and humidity	99% titre reduction (56h)	[106]
Steel	Omicron BA.1.1.529	$2.3 \times 10^4$ pfu	HCM/V/127	Room temperature and humidity	99% titre reduction (42h)	[106]
Laboratory equipment	SARS-CoV-2	5µL	Patient in the catarrhal phase	4 °C	t1/2: 273.36 h (glass slides)	[107]
Laboratory equipment	SARS-CoV-2	5µL	Patient in the catarrhal phase	24 °C	t1/2: 70.32 h (glass slides)	[107]
Non-porous surfaces	SARS-CoV-2	$4.97 \times 10^7$ (TCID50)	Betacoronavirus/Australia/SA01/2020	20 °C and 50%	t1/2: 1.68–2.74d;	[108]
Non-porous surfaces	SARS-CoV-2	$4.97 \times 10^7$ (TCID50)	Betacoronavirus/Australia/SA01/2020	30 °C and 50%	t1/2: 10.5–32.7 h	[108]
Non-porous surfaces	SARS-CoV-2	$4.97 \times 10^7$ (TCID50)	Betacoronavirus/Australia/SA01/2020	40 °C and 50%	t1/2: 1.4–3.0 h	[108]
Human-biological fluids	SARS-CoV-2	$1 \times 10^5$ (TCID50)	USA-WA1/2020	4°C and 40%	t1/2: 3.3 h and 5.8 h	[109]
Human-biological fluids	SARS-CoV-2	$1 \times 10^5$ (TCID50)	USA-WA1/2020	21 °C and 40%	t1/2: 3.1 h	[109]
Human-biological fluids	SARS-CoV-2	$1 \times 10^5$ (TCID50)	USA-WA1/2020	27 °C and 85%	t1/2: 1.5 h	[109]
Cat-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA/2020 strain	25 °C and 70%	t1/2: 2.84–12.74 h	[110]
Cat-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA/2020 strain	21 °C and 60%	t1/2: 2.71–8.1 h	[110]
Cat-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA/2020 strain	13 °C and 66%	t1/2: 7.42–18.38 h	[110]
Sheep-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA/2020 strain	25 °C and 70%	t1/2: 4.16–6.64 h	[110]
Sheep-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA/2020 strain	21 °C and 60%	t1/2: 6.10–9.33 h	[110]
Sheep-biological fluids	SARS-CoV-2	$5 \times 10^4$ (TCID50)	USA-WA/2020 strain	13 °C and 66%	t1/2: 9.04–99.95 h	[110]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID50)	HCoV-19 nCoV-WA1-2020	10 °C and 40%	t1/2: 26.55 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID50)	HCoV-19 nCoV-WA1-2020	10 °C and 65%	t1/2: 14.22 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID50)	HCoV-19 nCoV-WA1-2020	10 °C and 85%	t1/2: 13.78 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID50)	HCoV-19 nCoV-WA1-2020	22 °C and 40%	t1/2: 6.43 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID50)	HCoV-19 nCoV-WA1-2020	22 °C and 65%	t1/2: 2.41 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID50)	HCoV-19 nCoV-WA1-2020	22 °C and 85%	t1/2: 7.50 h	[111]

**Table 1** (continued)

Substrate	Virus	Virus titer	Type	Temperature (°C) and relative humidity (%)	Virus stability (hour (h))	Reference
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID <sub>50</sub> )	HCoV-19 nCoV-WA1-2020	27 °C and 40%	t1/2: 3.43 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID <sub>50</sub> )	HCoV-19 nCoV-WA1-2020	27 °C and 65%	t1/2: 1.52 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID <sub>50</sub> )	HCoV-19 nCoV-WA1-2020	27 °C and 85%	t1/2: 2.79 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID <sub>50</sub> )	HCoV-19 nCoV-WA1-2020	10 °C	t1/2: 42.08 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID <sub>50</sub> )	HCoV-19 nCoV-WA1-2020	22 °C	t1/2: 12.18 h	[111]
Polypropylene	SARS-CoV-2	$1 \times 10^5$ (TCID <sub>50</sub> )	HCoV-19 nCoV-WA1-2020	27 °C	t1/2: 5.76 h	[111]

Abbreviations: t1/2 Half-life, h hour, TCID<sub>50</sub> tissue culture infectious dose 50%/mL

table confirms that the virus is least active at 40% relative humidity. It was also found that temperature seemed to have a greater effect on viral activity than relative humidity. However, neither temperature nor humidity can instantly inactivate the virus under normal conditions (Table 1). Some controlled studies of human nasal mucus and sputum, as well as viral aerosols, have shown that SARS-CoV-2 decayed faster at higher relative humidity [101]. This is consistent with evidence of influenza virus survival that influenza is best transmitted at low absolute humidity [102, 103].

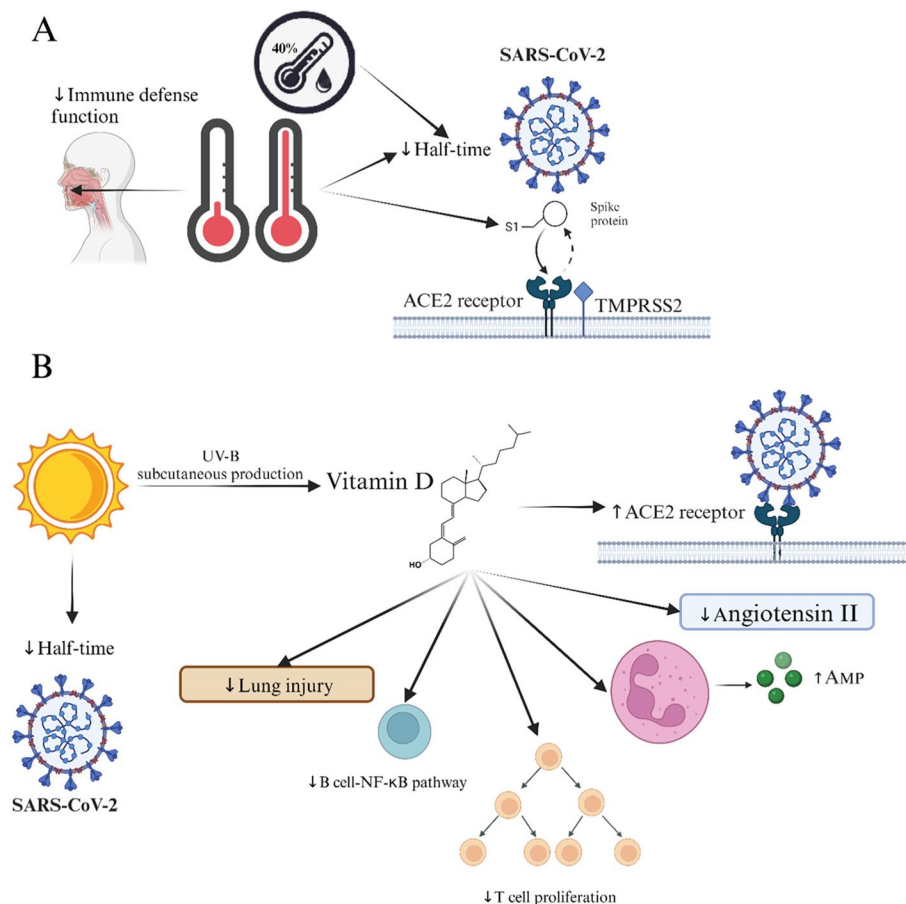
Some innate immune responses are suppressed at low temperatures. The human upper respiratory tract is the first contact site of inhaled respiratory viruses and the body's first line of defense against these foreign pathogens. Once bacteria are detected in the front of the nose, the epithelial cells of the anterior nasal mucosa increase the release of extracellular vesicles (EVs) several times. These EVs enter the nasal mucus and have a direct antibacterial effect. At the same time, EVs can arm the more rear epithelial cells with immunoprotective proteins, inducing a four-fold increase in the production of nitric oxide in epithelial cells [112]. In a subsequent study, Huang et al. further explored the EVs biological mechanism, but found that the EVs-mediated potent antiviral immune defense function was impaired by cold exposure. At ambient conditions of 4.4 °C, the number of EVs decreased by nearly 42%, while EVS-mediated functional delivery and the ability to neutralize viruses were weakened [113]. In addition, there is some research suggesting that cellular immune responses may also be affected by temperature and humidity. Mice airway epithelial cells initiated a stronger antiviral response at higher temperatures compared to lower temperatures [114], and mice exposed to low humidity conditions were more

susceptible to influenza infection [115]. However, the effects of seasonal fluctuations in immune response on COVID-19 susceptibility and severity are still largely unknown. When the temperature is low, the human immune response is suppressed and the activity of the virus is increased, which promotes the spread of the COVID-19. When the temperature is high, it will change the conformation of SARS-CoV-2 spike glycoprotein S1 and reduce the activity of the virus, thus inhibiting the transmission of COVID-19. At 40% RH, the activity of the virus is lowest, thus inhibiting the spread of the novel coronavirus (Fig. 1A).

#### Solar radiation

Among the different climatological factors, sunlight has been found to play an important role in determining the spread of SARS-CoV-2. Sunlight contains a spectrum of ultraviolet A (UVA), UVB, and UVC. UV germicidal is a commonly used disinfection method, and it has previously been reported that UV can inactivate aerosolized coronaviruses [116]. Lorca-Oro et al. used UV-C (100–280 nm wavelength) to inactivate SARS-CoV-2 in a laboratory simulating hospital intensive care unit conditions. The results showed that after 12 min or more of UV-C exposure, the titer was reduced by  $\geq 99.91\%$  to  $\geq 99.99\%$ , and the minimum distance between the UV-C device and the SARS-CoV-2 dry sample was 100 cm [117]. Under simulated sunlight conditions in the laboratory, the researchers found that 90% of SARS-CoV-2 was inactivated after 19 min of exposure under simulated winter and autumn UV conditions, while some degree of inactivation was achieved after just 8 min under simulated summer conditions [118]. Several other studies have found similar results, inactivating 90% of SARS-CoV-2 every 6.8 min in simulated saliva and every 14.3 min





**Fig. 1** Schematic diagram of the influence mechanism of natural environmental factors on the transmission of COVID-19. **A** Temperature. **B** Solar radiation. Promoting (upward arrow) or suppressing (downward arrow) the associated mechanism

in culture medium when exposed to simulated sunlight at the summer solstice (ultraviolet (UV) range: 280–400 nm) [119]. In the United States and most cities around the world, 90% or more of SARS-CoV-2 will be inactivated after 11–34 min of exposure to midday sunlight in the summer [120]. Observation of the above studies found that SARS-CoV-2 is inherently sensitive to UV. However, UVC can be absorbed by atmospheric ozone, and sunlight reaching the Earth's surface cannot directly eradicate SARS-CoV-2 through virus-killing activity [121].

UVB exposure is closely related to vitamin D synthesis. The body relies primarily on sun exposure to meet its vitamin D needs. UVB is absorbed by the 7-dehydrogenated cholesterol in the skin, causing it to be converted to pre-vitamin D3, which is quickly converted to vitamin D3 [122]. Significant increases in vitamin D can be achieved at very low UVB doses [123]. Solar radiation is highest in summer and at lower altitudes. Studies have found that in northern Europe, adequate vitamin D status can be achieved through summer sun exposure. In

winter, however, the UVB radiation in the environment is too low to produce any vitamin D [124]. Regardless of skin type and ethnicity, there is almost no vitamin D synthesis in winter and spring at latitudes  $>50^\circ$ . Vitamin D deficiency is associated with the severity of COVID-19. In a meta-analysis, vitamin D deficiency was found to be more severe in severe cases compared to mild cases. Insufficient vitamin D levels increase hospitalization rates and COVID-19 mortality [125]. Vitamin D can reduce the risk of COVID-19 in the following ways (Fig. 1B): (1) Vitamin D helps immune cells produce antimicrobial peptides, which play an antibacterial and antiviral role [126]; (2) Vitamin D can inhibit T cell proliferation and the NF- $\kappa$ B pathway of B cells, and reduce the level of pro-inflammatory cytokines [127]; (3) Vitamin D can prevent the constriction response of pulmonary blood vessels in COVID-19 [128]; (4) Vitamin D alleviates lung injury by stimulating endothelial cell proliferation and migration, reducing epithelial cell apoptosis, and inhibiting TGF- $\beta$ -induced epithelial-mesenchymal transformation [129]. More research is required to evaluate the mechanisms

whereby vitamin D might reduce the risk of COVID-19 [130].

### **Wind speed**

There is strong evidence to support airborne transmission of SARS-CoV-2 [131, 132]. The World Health Organization (WHO) has identified inhalation of virus-carrying aerosols as the primary mode of transmission of SARS-CoV-2 over short and long distances. Airborne transmission is defined as less than 5  $\mu\text{m}$  with >1 to 2 m from an infected person [133]. Wind speed can strongly influence the transport of virus-carrying aerosols [134]. Aerosols tend to rise because they are warmer than ambient air and are confined indoors by surrounding walls and ceilings [135]. In open spaces, particles or droplets produced by normal breathing can only be transferred over short distances, and when sneezing or coughing, particles carry nearly the same distance, with differences only within a certain range. The greater the air flow outside, the greater the dispersion. One study found that the distance of breathing particles is 0.65m, the distance of coughing is 1.63m and the distance of sneezing is 2.86m [136]. In addition, studies have assessed the risk of spreading infectious particulate matter while chamber musicians play their instruments. It turns out that no matter the volume, pitch, or content of the play, it did not extend the range of the air flow [137]. While wind speed cannot completely remove the spread of the virus, ventilation helps remove aerosols that carry the virus to reduce airborne transmission. For example, studies using simulations to track infected aerosol plumes in real time have found that a stable state of the atmosphere with low wind speeds, low-level turbulence, and cool, moist ground conditions facilitates the spread of disease. The trajectory model found that the virus can travel in the air for up to 30 min, covering a radius of 200 m at a time, 1–2 km away from the original source [138]. A study used computational fluid dynamics to simulate viral air flow in an office while investigating the effects of different ventilation strategies on viral transmission. The results showed that the ventilation strategy of single ventilation had the highest infection probability [139]. Another study found that ambient winds (wind velocities range from 0 to 16 km/h) increase the complexity of secondary flows. Even at 3.05 m, the droplets flow well along the air stream and deposit on the human body and head area. Due to wind convection, the remaining droplets can travel above 3.05 m in the air, posing a potential health risk to people nearby [140]. The study also found that a reduction in ventilation rates or room capacity per person, or an increase in the ratio of infected people to susceptible people, would increase the distance of transmission. Effective environmental prevention strategies

for respiratory infections require an appropriate increase in ventilation rates while maintaining sufficiently low occupancy rates [141]. Therefore, different ventilation strategies must be developed according to the actual indoor conditions to reduce the transmission of viruses in the air.

### **Environmental pollution factors**

#### **Particulate matter**

The relationship between air pollution and COVID-19 is well-established. Further research has found that air pollution can modify host susceptibility to infection and modify the severity of disease [142]. Table 2 summarizes some key in vivo studies of air pollution on COVID-19 related targets, immune cells, and oxidative stress. PM is the main component of air pollutants. Many studies have demonstrated that PM can increase the expression of angiotensin-converting enzyme 2 (ACE2) and transmembrane protease serine type 2 (TMPRSS2). SARS-CoV-2 can use ACE2 as an entry receptor and TMPRSS2 to activate S protein [143]. Cell studies have shown that ACE2 expression has become a risk factor for the development of COVID-19 [144]. Another study found that the expression of ACE2 and TMPRSS2 increased the infection rate of SARS-CoV-2 [145]. Therefore, PM can increase the expression of ACE2 and TMPRSS2 and affect the severity and incidence of COVID-19.

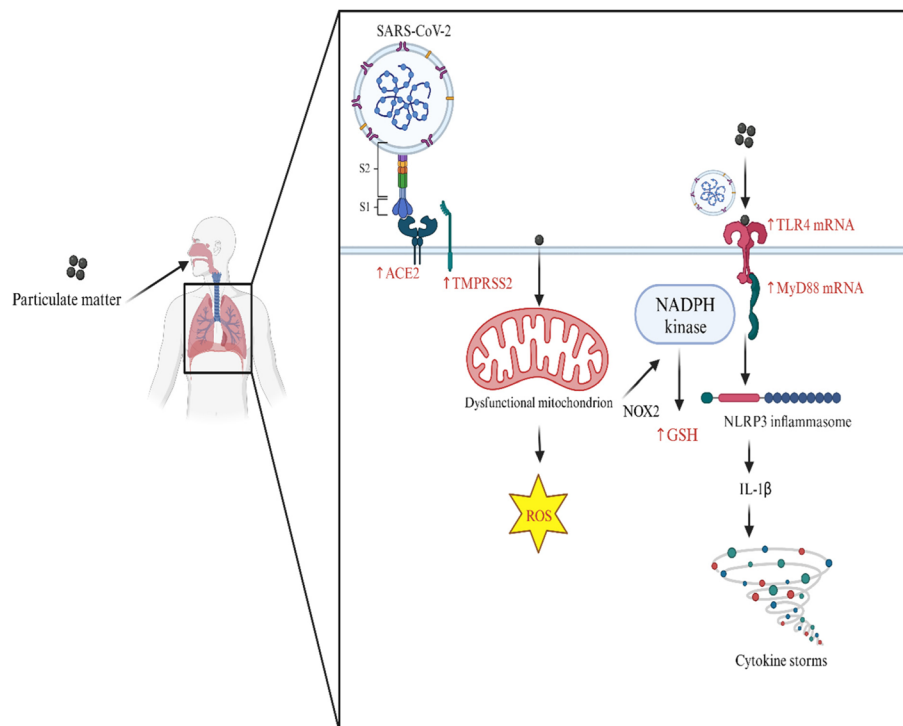
Severe COVID-19 is associated with high inflammation and elevated levels of inflammatory cytokines. Exposure to air pollutants increases the number of pro-inflammatory cytokines and immune cells that infiltrate the lungs, leading to systemic inflammation and immune disorders that reduce resistance to viruses (Fig. 2). Even low doses of PM<sub>2.5</sub> induce lung inflammation, oxidative stress, and worsening lung impedance and histology in mice [153]. Once pathogens establish themselves, inflammation of respiratory mucous membranes caused by exposure to air pollution may lead to a higher risk of severe COVID-19 outcomes through compound inflammation [155]. For example, studies in mouse models exposed to PM<sub>2.5</sub> have found that PM<sub>2.5</sub> may increase IL-1 $\beta$  secretion through the TLR4/MyD88 and NLRP3 inflammasome pathways, leading to airway inflammation in mice [149]. Intranasal transfer of pulmonary microbiota in PM<sub>2.5</sub> exposed mice has been found to influence PM<sub>2.5</sub> induced lung inflammation and oxidative stress, such as increased levels of pro-inflammatory cytokines and dysregulation of biomarkers associated with oxidative damage [154]. PM exposure may promote the development of cytokine storms in SARS-CoV-2 infection.

PM<sub>2.5</sub> not only damages the lungs directly exposed to air, but also causes pathological changes in other organ systems through excessive oxidative stress generated by

**Table 2** Mechanism research link air pollution to COVID-19

Model	Air pollutant dosage	Target	Effect	Reference
PM combined with BLM induced lung fibrosis mice (intratracheal administration)	200 µg/20 g	ACE2 and TMPRSS2	PM administration alone elevated the expression of ACE2 but not significantly; PM administration remarkably enhanced ACE2 and TMPRSS2 expression when combined with bleomycin treatment	[146]
PM <sub>2.5</sub> -treated mice (intratracheal instillation)	50 µl (500 µg/body)	ACE2 and TMPRSS2	PM-exposed lung tissues showed increased ACE2 and TMPRSS2 expression in the alveolar regions	[147]
PM <sub>1</sub> -treated rats	whole-body expose (3 and 6 months)	ACE2, AT1, and AT2	ACE2 had increased at 3 months and was significantly decreased at 6 months; smooth muscle hypertrophy had occurred after 6 months; AT1 and AT2 had significantly decreased at 6 months	[148]
PM <sub>2.5</sub> -treated mice (intratracheal instillation)	50 µl (0.5 mg)	IL-1β, TLR4/MyD88 pathway, and NLRP3 inflammasome	Induced acute lung inflammation by PM <sub>2.5</sub> exposure; ↑neutrophils, lymphocytes and macrophages; ↑TLR4, MyD88, NF-κB, NLRP3, P2X7 mRNA levels of lung tissues	[149]
PM <sub>2.5</sub> -treated mice (oral-nasal exposure system)	750 µg/m <sup>3</sup> (4h/6 weeks)	TNF-α, IL-1β, IL-18, HMGB1, NLRP3, ASC, and Caspase-1	↑TNF-α, IL-1β, and IL-18 levels; ↑mRNA expression levels of HMGB1, NLRP3, ASC, and Caspase-1 significantly	[150]
PM <sub>2.5</sub> -treated NC or HFC Apo E <sup>-/-</sup> mice	235.76 ± 72.73 µg/m <sup>3</sup> (8h/16 weeks)	TNF-α, NLRP3, ASC, caspase-1, IL-1β, and IL-18	Activated of NLRP3 inflammasome; ↓IL-10; ↑TNF-α, protein expression of NLRP3, ASC, caspase-1, IL-1β, and IL-18	[151]
Exposure to PM <sub>2.5</sub> in atmosphere and ambient aerosol-mice (C57BL/6, Nox2 <sup>-/-</sup> C57BL/6, Balb/c, Tlr4Lps-d (BALB/cAnPt), c-fmsYFP (FVB/N))	Facility (10.7 ± 2.1 µg/m <sup>3</sup> ) and chamber (92.4 ± 2.1 µg/m <sup>3</sup> ); 6 h/day 5 days/week	NADPH, TLR-dependent gene expression,	PM <sub>2.5</sub> increases inflammatory monocytes via TLR4 pathway; increase NADPH oxidase-derived superoxide; influence leukocyte trafficking in the microvasculature; ↑TLR-dependent gene expression; ↑TNFα, MCP-1, and IL12p70 in lung; ↓IL-10	[152]
PM <sub>2.5</sub> -treated BALB/c mice (intranasal instillation)	PM <sub>2.5</sub> from Sao Paulo, Brazil (10 µL or 30µL)	IL-6 and TNF-α	PM <sub>2.5</sub> increases elastic and viscoelastic components of lung mechanics; ↑MPO, IL-6, CAT, and TNF-α; neutrophil influx into lung parenchyma; ↓GSH/GSSG	[153]
PM <sub>2.5</sub> -treated ICR mice	3 days (25mg/kg)	Lung microbiota	↑TNF-α and IL-1β in BALF; ↓GSH, mt-Nd5, and mt-Nd6; ↑NPTX1, NLRP3; disrupted microbiota can influence inflammation and oxidative stress	[154]

**Abbreviations:** PM particulate matter, BLM bleomycin, AT1 angiotensin II receptor type, AT2 2, Ozone O<sub>3</sub>, IL-1β Interleukin-1 beta, TLR4 Toll-like receptor 4, MyD88 Myeloid differentiation primary response protein MyD88, NLRP3 LRR and PYD domains-containing protein 3, P2X7 P2X purinoceptor 7, IL-18 Interleukin-18, HMGB1 High mobility group protein B1, Apo E<sup>-/-</sup> apolipoprotein E<sup>-/-</sup>, ROS reactive oxygen species, Nr2f nuclear factor erythroid 2-related factor 2, TLR toll-like receptor



**Fig. 2** Schematic diagram of the influence of particulate matter on the transmission of COVID-19. Promote (up arrow, color red) related mechanisms

mitochondria [156]. Inflammation, oxidative stress, and cell death in alveolar epithelial cells caused increased mitochondrial division and decreased mitochondrial fusion when exposed to PM<sub>2.5</sub> [157]. The enzymes NOX2 (produce reactive oxygen species) and Toll-like receptor 4 (TLR), have been shown to be critical for PM-induced NADPH oxidase activation. PM<sub>2.5</sub> triggers an increase in phospholipid oxidation in the lungs, which then mediates systemic cellular inflammation through TLR4/NADPH oxidation-dependent mechanisms [152]. In the case of influenza A virus infection, activation of NOX2 oxidase can promote the production of reactive oxygen species to inhibit antiviral and humoral signaling networks [158]. Therefore, PM can increase the infection rate of SARS-CoV-2 by activating NOX2 to promote reactive oxygen species. Contaminant-induced oxidative stress and cell damage may worsen prognosis [159, 160]. Exposure to air pollution-induced oxidative stress is a key mechanism leading to cardiovascular morbidity and mortality [161].

### Ozone

Ozone is also a common air pollution in cities. It is a gaseous component that is produced by the interaction of air pollution components such as nitrogen oxides and organic compounds caused by sunlight. Inhaling ozone is very toxic to the lungs. Table 3 summarizes the

mechanisms by which ozone may be associated with COVID-19. After inhalation, ozone does not enter cells, but comes into direct contact with the first layer of cells on the surface of the airway, such as airway and alveolar epithelial cells and airway macrophages [162]. These cells release reactive oxygen species and various other inflammatory mediators, including cytokines and lipids, from oxidative damage to the airway epithelium [163]. Oxidative stress is a major pathogenic factor of COVID-19. For example, it has been found that ozone stimulated macrophages to secrete pro-inflammatory cytokines (IL-1 $\alpha$ , IL-1 $\beta$  and IL-18), and IL-1 $\alpha$  stimulated epithelial cells to secrete CXCL1 and CCL2, thereby driving neutrophil influx [164]. Another study found that canonical transient receptor potential 6 (TRPC6) regulates NF- $\kappa$ B activation and intercellular adhesion molecule-1 (ICAM-1) expression after exposure to ozone. TRPC6 deficiency attenuates O<sub>3</sub>-induced recruitment of neutrophils to airway epithelial cells and ICAM-1 expression [165]. In addition, ozone can lead to loss of antioxidant Nrf2 and SOD activity in the body, enhanced intracellular oxidative stress and increased HIF-1 $\alpha$  signaling, resulting in a persistent chronic inflammatory environment in the lungs [166]. Ozone can induce the expression of MAPK, NF- $\kappa$ B and AP-1 proteins through TLR4/MyD88 pathway, resulting in inflammatory response. Heat shock protein 70



**Table 3** Mechanism research link ozone to COVID-19

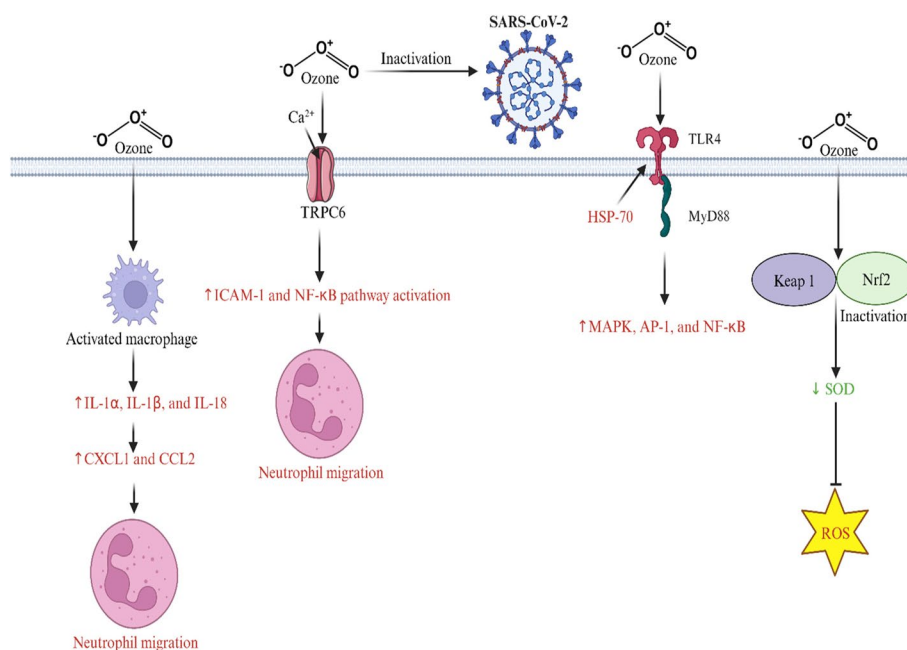
Model	Air pollutant dosage	Target	Effect	Reference
O3-treated OVA induced asthma mice	3 ppm mixed with air for 3 h (expose one hour)	ROS	O3 exposure increased ROS release and asthma exacerbation and elevated neutrophil lung infiltration; O3 increased pro-inflammatory cytokine production as well as the percentage of IL-17A <sup>+</sup> γδT cells	[170]
O3-treated rat alveolar macrophage cell line NR8383; NR8383-conditioned media-treated alveolar type I-like cells	100, 200, and 400 ppb for 1 h 24 h in media	IL-1α, IL-1β, CXCL1, CCL2	↑IL-1α, IL-1β, GM-CSF, and IL-18 in supernatant of NR8383 cells exposed to 100 ppb ozone; ↑CINC-1 and MCP-1 in supernatants from the epithelial cells;	[164]
O3-treated wild-type, TRPC6 <sup>-/-</sup> , and TRPC6 <sup>+/+</sup> C57BL/6 mice; human airway epithelial cell (human airway epithelial cell)	1 ppm for 3 h/day with repeated exposure every other day (days 1, 3, and 5)	TRPC6	O3-induced neutrophil recruitment to airway epithelial cells; ↑ICAM-1; TRPC6-dependent Ca <sup>2+</sup> entry ICAM-1-mediated O3-induced neutrophil adhesion to airway epithelial cells	[165]
O3-treated C57BL/6 mice	3 ppm for 3 h/day, 2/week (1, 3 or 6 weeks)	HDAC2, Nrf2, and HIF-1α	Induce lung injury; increase inflammatory cell number; ↑KC, IFN-γ, MIF, TNF-α, IL-6, and IL-17; ↑SOD, HIF-1α, Nrf2, Gpx1 mRNA, Txnrd1 mRNA, pHDAC2; ↓HDAC2	[166]
O3-treated C3H/HeJ (HeJ; Tlr4 mutant) and C3H/HeOuJ (OuJ; Tlr4 normal) mice	0.3 ppm for 6, 24, 48, or 72 h	TLR4	↑p65, AP-1; ↑transcript levels of Myd88 and Trif; ↑ERK1/2 and p38;	[167]
O3-treated human (13 healthy, 4 allergic non asthmatics, 10 allergic asthmatics)	0.4 ppm for 2 h	IL-1β and IL-8	Post exposure, show activated innate immune function; ↑IL-1β and IL-8;	[171]
O3-treated OVA-established asthmatic mouse model	1 ppm for 3 h/day/3 times	ROS	Enhance AHR and decrease CL; increase the number of total leukocytes, macrophages, neutrophils; ↑HA, IFN-γ, CXCL-1, TNF-α, and IL-17; ↑phosphorylation level of p38 MAPK and HSP27; ↑MDA, GSH-Px;	[172]

**Abbreviations:** h hour, CXCL1 cytokine-induced neutrophil chemoattractant-1, CCL2 monocyte chemoattractant protein-1, TRPC6 canonical transient receptor potential 6, ICAM-1 intercellular adhesion molecule-1, Gpx1 glutathione peroxidase 1, HDAC2 histone deacetylase 2, Nrf2 nuclear factor erythroid-related factor 2, HIF-1α hypoxia-inducible factor-1α, ERK1/2 extracellular-signal-related kinase-1/2, AP-1 activator protein-1, AHR airway hyperresponsiveness, CL lung compliance, HA hyaluronan

(HSP70) was identified as a downstream mediator with ozone mediated TLR4 effects [167]. Ozone also induced apoptosis markers (lysed caspase 9) and autophagy markers (beclin-1) in alveolar macrophages and enhanced the expression of MMP-2 and MMP-9 [168]. However, the effects of ozone on different cell death pathways such as necrosis, apoptosis, ferroptosis, and autophagy have not been resolved [169]. These cell death pathways may be responsible for the emphysema process induced by oxidants. Just as ozone can make asthma worse, ozone may increase the incidence and severity of COVID-19 by inducing inflammation, oxidative stress, and airway remodeling (Fig. 3).

Ozone is an excellent biocidal agent due to its strong oxidation, and its effectiveness against bacteria, fungi and viruses has been proven [173]. Ozone can be easily applied to large and small areas for disinfection and is broken down back into safe oxygen after treatment. Ozone is particularly deadly to viruses through peroxidation of lipid surface and subsequent damage to lipid envelopes and proteins, and enveloped viruses such as SARS-CoV-2 are more vulnerable to ozone attack [174]. Ozone has been shown to inactivate the SARS-CoV-2 virus on surfaces (such as plastic, glass, stainless steel, gauze, wood, wool, copper, and coupons in ambulance seats and floors) or in suspended fluids [175]. During the pandemic, ozone has been widely used to purify many enclosed spaces.

Ozone also has medical uses. Medical ozone is administered in the form of a balanced O<sub>2</sub>/O<sub>3</sub> mixture by autologous blood therapy or rectal blow or also as a peritoneal injection in laboratory animals [176]. Medical ozone can interfere with the replication phase of the virus to play an antiviral role. Medical ozone's effects include the oxidation and inactivation of specific viral receptors used to form cell-membrane binding structures, thereby inhibiting the level of its first stage: cellular penetration [177]. Medical ozone can directly act on Nrf2, an important nuclear message transmitter, regulating and blocking the activity of ACE2 receptors. Thus, preventing SARS-CoV-2 from replicating [178]. SARS-CoV2 can cause oxidative stress and inflammation, further tissue damage and widespread triggering of the clotting cascade, culminating in the formation of blood clots [179]. Medical ozone, when administered in appropriate pathways and at small doses, may induce adaptive responses that reduce endogenous oxidative stress [180]. In addition, some studies have found that ozone can activate the cellular and humoral immune system and can reduce inflammation/apoptosis processes [177]. Clinical studies have further confirmed that ozone therapy can be used as a comprehensive treatment for COVID-19 with low cost and improve the health status of patients [181].



**Fig. 3** Schematic diagram of the influence of ozone on the transmission of COVID-19. Promote (up arrow, color red) or suppressing (downward arrow, color green) related mechanisms

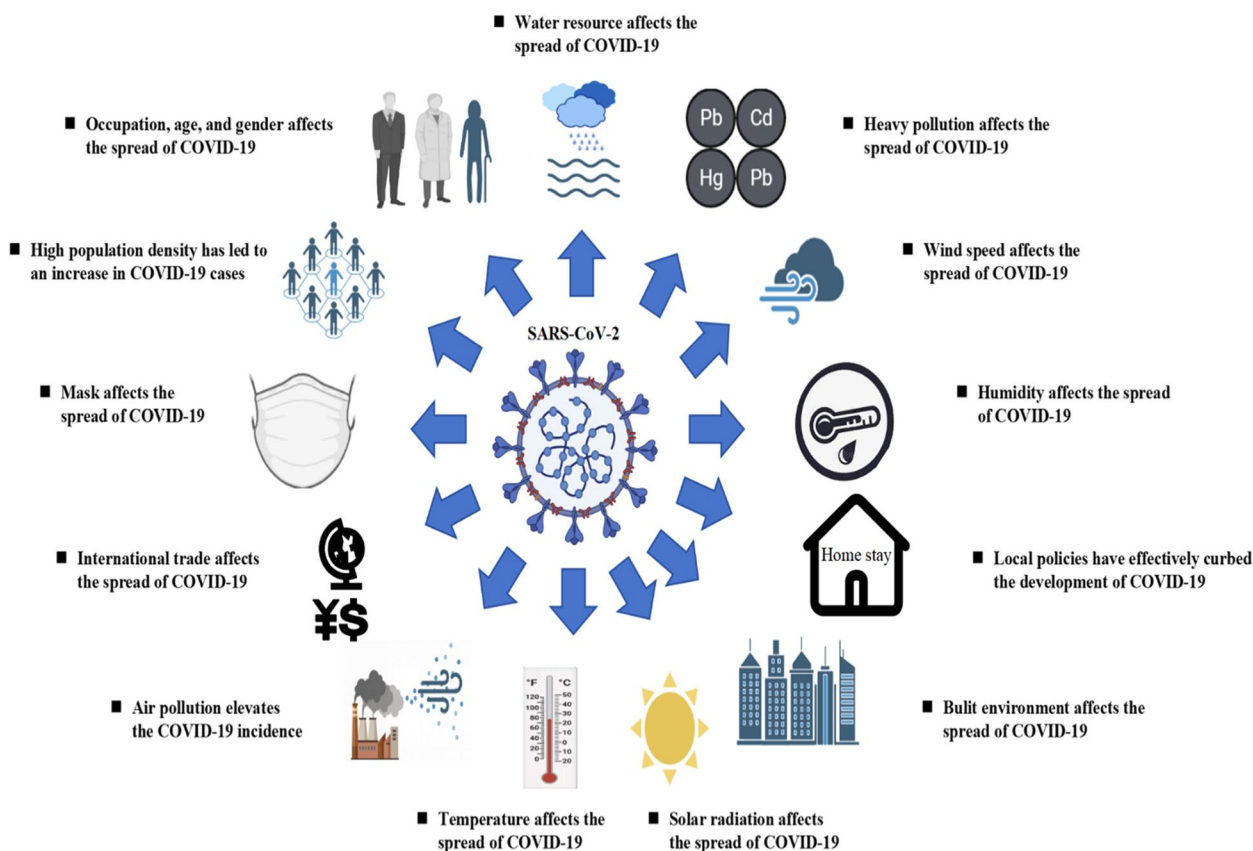
### Discussion

Since the outbreak, extensive research has investigated the factors that influence the spread of COVID-19. Representative studies on the impact of natural environmental factors and social environmental factors on COVID-19 are discussed in this review. The mechanisms and results of natural environmental factors affecting COVID-19 are shown in Fig. 4. As can be seen from the epidemiological studies included in this paper, the current study covered multiple regions, multiple confounding factors, and long or short-term exposure times. The mechanism study also covers multiple pathways and targets in vivo and in vitro.

Both mechanism and epidemiological studies have shown that air pollution, especially PM2.5 and ozone, greatly accelerates the spread of COVID-19. Some of the differences in air pollution effectiveness in transmitting the virus may be caused by differences in composition between different locations. However, there are important limitations to the available evidence, such as (1) methodological limitations, (2) incomplete coverage of

the original data, and (3) large uncertainties in the analysis [182, 183]. Studies on the mechanism of air pollution affecting the spread of COVID-19 have confirmed that air pollution can (1) increase the expression of key proteins in the entry pathway of SARS-CoV-2, (2) promote inflammation and release of pro-inflammatory cytokines, (3) causes pathological changes in organ systems, and (4) increase the risk of respiratory complications. This further provides evidence for COVID-19 prevention and control measures to reduce air pollution, rational use of ozone disinfection, and medical ozone therapy. To date, no study has accurately demonstrated seasonal changes in the global prevalence of COVID-19. Research on climate conditions is subject to similar challenges and limitations as air pollution [184]. But a growing body of evidence supports a statistically significant correlation between climatic conditions and morbidity, mortality, recovery cases, etc. [185]. In vitro experiments were conducted to study the activity of the virus under different climatic conditions and substrate conditions. Studies have shown that the SARS-CoV-2 has the longest half-life

### Natural and social environmental factors and transmission of COVID-19



**Fig. 4** Schematic diagram of nature and social environment factors affects the transmission of COVID-19

at lower temperatures, which promotes SARS-CoV-2 transmission. This also provides evidence for COVID-19 prevention and control measures to maintain indoor temperature and humidity. SARS-CoV-2 rots faster at higher relative humidity, inhibiting the spread of the virus. Solar radiation has a better inactivation effect on SARS-CoV-2, but UVC in the environment cannot directly eradicate SARS-CoV-2. Solar radiation (UVB) inhibits the spread and development of COVID-19 through the synthesis of vitamin D. Therefore, there is scientific evidence for COVID-19 prevention and control measures based on rational exposure to sunlight. For other climatic conditions (such as wind speed, water resource), no clear conclusions have been found.

Social environmental factors are also significantly related to the spread of COVID-19. The mechanisms and results of social environmental factors affecting COVID-19 are shown in Fig. 4. People vary greatly in their daily routines, traveling from home, work, school, and public and commercial spaces can exhibit high personal exposure to pathogens [186]. Epidemiologic studies have shown that population density, built environment, occupation, age, gender, local policies and socioeconomic activity have an impact on COVID-19 [187]. This is also the scientific basis for the country to take measures such as control of population movement, control of population density, wearing masks, and disinfecting dense places [188]. Among them, population density is considered to have a more significant impact on COVID-19 than meteorological factors, and the increase in population density will in turn lead to an increase in COVID-19 cases and deaths [73]. In addition, socio-economic activities such as international trade are also key factors affecting the spread of COVID-19. The total import and export volume is highly positively correlated with confirmed cases [97]. Although there are differences between the results of the studies, this may be due to the limitations of the studies introducing bias. Like the limitations described earlier, single factor studies can be biased due to confounding factors at the individual level.

### Future prospects

Now, as vaccination coverage increases and strains mutate, COVID-19 may be transitioning to an epidemic seasonal disease, such as influenza [189]. Climate may play a bigger role in determining COVID-19 infection. In the near term, as public health measures are reduced, the link between COVID-19 and natural environmental factors will become clearer. Future studies are needed to determine the effects of climate change on the spatiotemporal distribution of different strains of viruses. In addition, future research could focus on disease outcomes caused by climatic conditions in animal models

of COVID-19, and further improve the range of environmental conditions used in laboratory studies to better simulate real-world environmental conditions (indoor and outdoor). A meta-analysis of climate-related epidemiologic should be attempted to provide more conclusive evidence.

In addition, it is worth studying whether long-term environmental changes and short-term climate changes have the same effect on the human body. The impact of environmental exposure changes throughout life [190]. We do not know how the duration of exposure will affect the susceptibility and severity of COVID-19. Existing mechanism research is focused on animal experiments, and ethical clinical trials are needed. Existing mechanism studies have found that both temperature and humidity, as well as air pollution, can have an impact on immunity. Future research should address the interaction between climate and immunity. Explore the specific mechanism of climate on immunity through clinical research.

### Conclusions

The impact of COVID-19 on human health is significantly negative. The constant change of the current environment increases the probability of infectious diseases. Sorting out the key factors affecting infectious diseases for scientific prevention and control, personalized and precise treatment is critical, although there is still a lot of work to be done. Our review indicates that with the continuous mutation of SARS-CoV-2, high temperature, high humidity, low air pollution levels, and low population density may be more likely to slow down the spread of the virus. All of these measures appear to be effective in controlling the spread or mortality of COVID-19: (1) reducing air pollution levels, (2) rational use of ozone disinfection and medical ozone therapy, (3) rational exposure to sunlight, (4) scientific ventilation and maintenance of indoor temperature and humidity, (5) control of population density, and (6) control of population movement. They could play a vital role in the future face of infectious diseases. The arrival of new pathogens is inevitable. While focusing on the research and development of vaccines, diagnostic reagents, and drugs for infectious diseases, we use interdisciplinary methods to break through existing limitations and clarify the impact of environment on biology, disease and evolution from the molecular level with the development of methodology. It is an urgent need to safeguard people's health. This will provide innovative inspiration for global epidemic prevention and control policies and provide reference for similar infectious diseases that may emerge in the future.

### Acknowledgements

We thank BioRender (Gallery (biorender.com)) for its online mapping services provided during the writing of this manuscript.



**Authors' contributions**

Conceptualization: Yanping Wang, Bin Liu, and Nannan Shi; original draft preparation, Zhaoyuan Gong, Tian Song, and Mingzhi Hu; writing-review & editing, Qianzi Che, Jing Guo; and project administration, Haili Zhang and Huizhen Li. All authors read and approved the final manuscript.

**Funding**

This work was supported by the China Academy of Chinese Medical Sciences key collaborative project of Innovation Fund, Analysis of Dynamic Temporal and Spatial Characteristics of TCM Syndrome of COVID-19 and Research on Its Biological Connotation, (No.CI2022C004-01), National Key R&D Program of China (2023YFC3503400), the 2023 Agency/National Special Fund Project of National Administration of Traditional Chinese Medicine (F0212), and 2022 Qi Huang Young Scholar programme of the National Administration of Traditional Chinese Medicine (Nannan Shi) (Z0841, Z0865).

**Availability of data and materials**

Not applicable.

**Data availability**

No datasets were generated or analysed during the current study.

**Declarations****Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare no competing interests.

**Author details**

<sup>1</sup>Institute of Basic Research in Clinical Medicine, China Academy of Chinese Medical Sciences, Beijing 100700, China.

Received: 22 January 2024 Accepted: 9 August 2024

Published online: 13 August 2024

**References**

- Kamacooko O, Kitonsa J, Bahemuka UM, Kibengo FM, Wajja A, Basajja V, et al. Knowledge, attitudes, and practices regarding covid-19 among healthcare workers in uganda: a cross-sectional survey. *Int J Environ Res Public Health*. 2021;18:7004.
- Sarhan RM, Madney YM, Abou Warda AE, Boshra MS. Therapeutic efficacy, mechanical ventilation, length of hospital stay, and mortality rate in severe COVID-19 patients treated with tocilizumab. *Int J Clin Pract*. 2021;75:e14079.
- Sarhan RM, Mohammad MF, Boshra MS. Differential clinical diagnosis and prevalence rate of allergic rhinitis, asthma and chronic obstructive pulmonary disease among COVID-19 patients. *Int J Clin Pract*. 2021;75:4–6.
- Barouki R, Kogevinas M, Audouze K, Belesova K, Bergman A, Birnbaum L, et al. The COVID-19 pandemic and global environmental change: emerging research needs. *Environ Int*. 2021;146:106272.
- Li M, Wang H, Tian L, Pang Z, Yang Q, Huang T, et al. COVID-19 vaccine development: milestones, lessons and prospects. *Signal Transduction and Targeted Therapy*. 2022;7(1):146.
- Xiao S, Qi H, Ward MP, Wang W, Zhang J, Chen Y, et al. Meteorological conditions are heterogeneous factors for COVID-19 risk in China. *Environ Res*. 2020;2021(198):111182.
- Scannell Bryan M, Sun J, Jagai J, Horton DE, Montgomery A, Sargis R, et al. Coronavirus disease 2019 (COVID-19) mortality and neighborhood characteristics in Chicago. *Ann Epidemiol*. 2021;56:47–54.e5.
- Weaver AK, Head JR, Gould CF, Carlton EJ, Remais JV. Environmental Factors Influencing COVID-19 Incidence and Severity. *Annu Rev Public Health*. 2022;43:271–91.
- Weaver AK, Head JR, Gould CF, Carlton EJ, Remais JV. Environmental factors influencing COVID-19 incidence and severity. 2022. <https://doi.org/10.1146/annurev-publhealth>.
- Huang Z, Huang J, Gu Q, Du P, Liang H, Dong Q. Optimal temperature zone for the dispersal of COVID-19. *Sci Total Environ*. 2020;736:139487.
- Azuma K, Kagi N, Kim H, Hayashi M. Impact of climate and ambient air pollution on the epidemic growth during COVID-19 outbreak in Japan. *Environ Res*. 2020;190:110042.
- Haque SE, Rahman M. Association between temperature, humidity, and COVID-19 outbreaks in Bangladesh. *Environ Sci Policy*. 2020;114:253–5.
- Tan J, Mu L, Huang J, Yu S, Chen B, Yin J. An initial investigation of the association between the SARS outbreak and weather: with the view of the environmental temperature and its variation. *J Epidemiol Community Health*. 1978;2005(59):186–92.
- Yin C, Zhao W, Pereira P. Meteorological factors' effects on COVID-19 show seasonality and spatiality in Brazil. *Environ Res*. 2021;2022(208):112690.
- Mecenas P, da Rosa Moreira Bastos RT, Rosário Vallinoto AC, Normando D. Effects of temperature and humidity on the spread of COVID-19: a systematic review. *PLoS One*. 2020;15(9):e0238339.
- Zheng HL, Guo ZL, Wang ML, Yang C, An SY, Wu W. Effects of climate variables on the transmission of COVID-19: a systematic review of 62 ecological studies. *Environ Sci Pollut Res*. 2021;28:54299–316.
- Yuan J, Wu Y, Jing W, Liu J, Du M, Wang Y, et al. Association between meteorological factors and daily new cases of COVID-19 in 188 countries: A time series analysis. *Sci Total Environ*. 2021;780:146538.
- Yuan J, Wu Y, Jing W, Liu J, Du M, Wang Y, et al. Non-linear correlation between daily new cases of COVID-19 and meteorological factors in 127 countries. *Environ Res*. 2021;193:110521.
- Mehta SK, Ananthavel A, Reddy TVR, Ali S, Mehta SB, Kakkanattu SP, et al. Indirect Response of the Temperature, Humidity, and Rainfall on the Spread of COVID-19 over the Indian Monsoon Region. *Pure Appl Geophys*. 2023;180:383–404.
- Gupta A, Banerjee S, Das S. Significance of geographical factors to the COVID-19 outbreak in India. *Model Earth Syst Environ*. 2020;6:2645–53.
- Cai QC, Lu J, Xu QF, Guo Q, Xu DZ, Sun QW, et al. Influence of meteorological factors and air pollution on the outbreak of severe acute respiratory syndrome. *Public Health*. 2007;121:258–65.
- Berendt RF, Dorsey EL. Effect of simulated solar radiation and sodium fluorescein on the recovery of Venezuelan equine encephalomyelitis virus from aerosols. *Appl Microbiol*. 1971;21:447–50.
- Rosario DKA, Mutz YS, Bernardes PC, Conte-Junior CA. Relationship between COVID-19 and weather: case study in a tropical country. *Int J Hyg Environ Health*. 2020;229:113587.
- Ahmadi M, Sharifi A, Dorosti S, Jafarzadeh Ghouschi S, Ghanbari N. Investigation of effective climatology parameters on COVID-19 outbreak in Iran. *Sci Total Environ*. 2020;729:138705.
- Lansiaux É, Pébay PP, Picard JL, Forget J. Covid-19 and vit-d: disease mortality negatively correlates with sunlight exposure. *Spat Spatiotemporal Epidemiol*. 2020;35:100362.
- Hashim BM, Al-Naseri SK, Hamadi AM, Mahmood TA, Halder B, Shahid S, et al. Seasonal correlation of meteorological parameters and PM2.5 with the COVID-19 confirmed cases and deaths in Baghdad, Iraq. *Int J Disaster Risk Reduct*. 2023;94:103799.
- Asyary A, Veruswati M. Sunlight exposure increased Covid-19 recovery rates: a study in the central pandemic area of Indonesia. *Sci Total Environ*. 2020;729:139016.
- Al-Khateeb MS, Abdulla FA, Al-Delaimy WK. Long-term spatiotemporal analysis of the climate related impact on the transmission rate of COVID-19. *Environ Res*. 2023;236:116741.
- Sharun K, Tiwari R, Dhama K. COVID-19 and sunlight: Impact on SARS-CoV-2 transmissibility, morbidity, and mortality. *Ann Med Surg*. 2021;66:17–20.
- Wang J, Li W, Yang B, Cheng X, Tian Z, Guo H. Impact of hydrological factors on the dynamic of COVID-19 epidemic: a multi-region study in China. *Environ Res*. 2021;198:110474.

31. Tosepu R, Gunawan J, Effendy DS, Ahmad LOAI, Lestari H, Bahar H, et al. Correlation between weather and Covid-19 pandemic in Jakarta, Indonesia. *Sci Total Environ.* 2020;725:138436.
32. Bashir MF, Ma B, Bilal, Komal B, Bashir MA, Tan D, et al. Correlation between climate indicators and COVID-19 pandemic in New York, USA. *Sci Total Environ.* 2020;728:138835.
33. Auler AC, Cássaro FAM, da Silva VO, Pires LF. Evidence that high temperatures and intermediate relative humidity might favor the spread of COVID-19 in tropical climate: a case study for the most affected Brazilian cities. *Sci Total Environ.* 2020;729:139090.
34. Domingo JL, Rovira J. Effects of air pollutants on the transmission and severity of respiratory viral infections. *Environ Res.* 2020;187:109650.
35. Copat C, Cristaldi A, Fiore M, Grasso A, Zuccarello P, Signorelli SS, et al. The role of air pollution (PM and NO<sub>2</sub>) in COVID-19 spread and lethality: a systematic review. *Environ Res.* 2020;191:110129.
36. Wu X, Nethery RC, Sabath MB, Braun D, Dominici F. Air pollution and COVID-19 mortality in the United States: strengths and limitations of an ecological regression analysis. *Sci Adv.* 2020;6:1–7.
37. Bossak BH, Andritsch S. COVID-19 and Air pollution: a spatial analysis of particulate matter concentration and pandemic-associated mortality in the US. *Int J Environ Res Public Health.* 2022;19:592.
38. Coker ES, Cavalli L, Fabrizi E, Guastella G, Lippo E, Parisi ML, et al. The effects of air pollution on COVID-19 related mortality in Northern Italy. *Environ Resour Econ (Dordr).* 2020;76:611–34.
39. Cole MA, Ozgen C, Strobl E. Air pollution exposure and Covid-19 in Dutch Municipalities. *Environ Resour Econ (Dordr).* 2020;76:581–610.
40. Travaglio M, Yu Y, Popovic R, Selley L, Leal NS, Martins LM. Links between air pollution and COVID-19 in England. *Environ Pollut.* 2021;268:115859.
41. Zhang X, Tang M, Guo F, Wei F, Yu Z, Gao K, et al. Associations between air pollution and COVID-19 epidemic during quarantine period in China. *Environ Pollut.* 2021;268:115897.
42. Zhou X, Josey K, Kamareddine L, Caine MC, Liu T, Micklely LJ, et al. Excess of COVID-19 cases and deaths due to fine particulate matter exposure during the 2020 wildfires in the United States. *Sci Adv.* 2021;7:1–12.
43. Lavigne E, Rytli N, Gasparri A, Sera F, Weichenthal S, Chen H, et al. Short-term exposure to ambient air pollution and individual emergency department visits for COVID-19: A case-crossover study in Canada. *Thorax.* 2023;78:459–66.
44. Liu Z, Liang Q, Liao H, Yang W, Lu C. Effects of short-term and long-term exposure to ambient air pollution and temperature on long recovery duration in COVID-19 patients. *Environ Res.* 2023;216:114781.
45. Ponedzialek B, Rzymyski P, Zarębska-Michaluk D, Rogalska M, Rorat M, Czupryna P, et al. Short-term exposure to ambient air pollution and COVID-19 severity during SARS-CoV-2 delta and omicron waves: a multicenter study. *J Med Virol.* 2023;95:e28962.
46. Vos S, De Waele E, Goeminne P, Bijmens EM, Bongaerts E, Martens DS, et al. Pre-admission ambient air pollution and blood soot particles predict hospitalisation outcomes in COVID-19 patients. *Eur Respir J.* 2023;62:2300309.
47. Alaniz AJ, Carvajal MA, Carvajal JG, Vergara PM. Effects of air pollution and weather on the initial COVID-19 outbreaks in United States, Italy, Spain, and China: a comparative study. *Risk Anal.* 2023;43:8–18.
48. Sidell MA, Chen Z, Huang BZ, Chow T, Eckel SP, Martinez MP, et al. Ambient air pollution and COVID-19 incidence during four 2020–2021 case surges. *Environ Res.* 2022;208:112758.
49. Lipsitt J, Chan-Golston AM, Liu J, Su J, Zhu Y, Jerrett M. Spatial analysis of COVID-19 and traffic-related air pollution in Los Angeles. *Environ Int. D* 2020;2021(153):106531.
50. Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, et al. First Case of 2019 Novel Coronavirus in the United States. *N Engl J Med.* 2020;382:929–36.
51. Peiris JSM, Chu CM, Cheng VCC, Chan KS, Hung IFN, Poon LLM, et al. Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. *Lancet.* 2003;361:1767–72.
52. Izquierdo-Lara R, Elsinga G, Heijnen L, Oude Munnink BB, Schapendonk CME, Nieuwenhuijse D, et al. Monitoring SARS-CoV-2 circulation and diversity through community wastewater sequencing, the Netherlands and Belgium. *Emerg Infect Dis.* 2021;27:1405–15.
53. Ahmed W, Angel N, Edson J, Bibby K, Bivins A, O'Brien JW, et al. First confirmed detection of SARS-CoV-2 in untreated wastewater in Australia: a proof of concept for the wastewater surveillance of COVID-19 in the community. *Sci Total Environ.* 2020;728:138764.
54. Wu F, Zhang J, Xiao A, Gu X, Lee L, Armas F, et al. SARS-CoV-2 titers in wastewater are higher than expected. *mSystems.* 2020;5:1–9.
55. Carraturo F, Del Giudice C, Morelli M, Cerullo V, Libralato G, Galdiero E, et al. Persistence of SARS-CoV-2 in the environment and COVID-19 transmission risk from environmental matrices and surfaces. *Environ Pollut.* 2020;265:115010.
56. Rimoldi SG, Stefani F, Gigantiello A, Polesello S, Comandatore F, Mileto D, et al. Presence and infectivity of SARS-CoV-2 virus in wastewaters and rivers. *Sci Total Environ.* 2020;744:140911.
57. Rahman MM, Bodrud-Doza M, Griffiths MD, Mamun MA. Biomedical waste amid COVID-19: perspectives from Bangladesh. *Lancet Glob Health.* 2020;8:e1262.
58. Martínez-Puchol S, Rusiñol M, Fernández-Cassi X, Timonedá N, Itarte M, Andrés C, et al. Characterisation of the sewage virome: comparison of NGS tools and occurrence of significant pathogens. *Sci Total Environ.* 2020;713:136604.
59. Wiktorczyk-Kapischke N, Grudlewska-Buda K, Walecka-Zacharska E, Kwiecińska-Piróg J, Radtke L, Gospoderek-Komkowska E, et al. SARS-CoV-2 in the environment—non-droplet spreading routes. *Sci Total Environ.* 2021;770:85–94.
60. Anand U, Bianco F, Suresh S, Tripathi V, Núñez-Delgado A, Race M. SARS-CoV-2 and other viruses in soil: an environmental outlook. *Environ Res.* 2021;198:111297.
61. Gong Z, Chan HT, Chen Q, Chen H. Application of nanotechnology in analysis and removal of heavy metals in food and water resources. *Nanomaterials.* 2021;11:1–32.
62. Park SK, Sack C, Sirén MJ, Hu H. Environmental cadmium and mortality from influenza and pneumonia in U.S. adults. *Environ Health Perspect.* 2020;128:127004-1-127004-8.
63. Skalny AV, Lima TRR, Ke T, Zhou JC, Bornhorst J, Alekseenko SI, et al. Toxic metal exposure as a possible risk factor for COVID-19 and other respiratory infectious diseases. *Food Chem Toxicol.* 2020;146:111809.
64. Hu X, Kim KH, Lee Y, Fernandes J, Smith MR, Jung YJ, et al. Environmental cadmium enhances lung injury by respiratory syncytial virus infection. *Am J Pathol.* 2019;189:1513–25.
65. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* 2020;395:1033–4.
66. Christenson SA, Smith BM, Bafadhel M, Putcha N. Chronic obstructive pulmonary disease. *Lancet.* 2022;399:2227–42.
67. Solenkova NV, Newman JD, Berger JS, Thurston G, Hochman JS, Lamas GA. Metal pollutants and cardiovascular disease: mechanisms and consequences of exposure. *Am Heart J.* 2014;168:812–22.
68. Wang X, Mukherjee B, Park SK. Associations of cumulative exposure to heavy metal mixtures with obesity and its comorbidities among U.S. adults in NHANES 2003–2014. *Environ Int.* 2018;121:683–94.
69. Sattar N, McInnes IB, McMurray JJV. Obesity Is a Risk Factor for Severe COVID-19 Infection: Multiple Potential Mechanisms. *Circulation.* 2020;142:4–6.
70. Zeng HL, Yang Q, Yuan P, Wang X, Cheng L. Associations of essential and toxic metals/metalloids in whole blood with both disease severity and mortality in patients with COVID-19. *FASEB J.* 2021;35:e21392.
71. Zeng HL, Zhang B, Wang X, Yang Q, Cheng L. Urinary trace elements in association with disease severity and outcome in patients with COVID-19. *Environ Res. D* 2020;2021(194):110670.
72. Zhang X, Maggioni V, Houser P, Xue Y, Mei Y. The impact of weather condition and social activity on COVID-19 transmission in the United States. *J Environ Manage.* 2022;302(Pt B):114085.
73. Ganasegeran K, Jamil MFA, Ch'ng ASH, Looi I, Peariasamy KM. Influence of population density for COVID-19 spread in Malaysia: an ecological study. *Int J Environ Res Public Health.* 2021;18:9866.
74. Wong HS, Hasan MZ, Sharif O, Rahman A. Effect of total population, population density and weighted population density on the spread of Covid-19 in Malaysia. *PLoS One.* 2023;18(4):1–15.
75. Md Iderus NH, Lakha Singh SS, Mohd Ghazali S, Yoon Ling C, Cia Vei T, Md Zamri ASS, et al. Correlation between population density and COVID-19 cases during the third wave in Malaysia: effect of the delta variant. *Int J Environ Res Public Health.* 2022;19:7439.

76. Diao Y, Koderá S, Anzai D, Gomez-Tames J, Rashed EA, Hirata A. Influence of population density, temperature, and absolute humidity on spread and decay durations of COVID-19: a comparative study of scenarios in China, England, Germany, and Japan. *One Health*. 2020;2021(12):100203.
77. Pinter-Wollman N, Jelic A, Wells NM. The impact of the built environment on health behaviours and disease transmission in social systems. *Biological Sciences: Philosophical Transactions of the Royal Society B*; 2018. p. 373.
78. Kan Z, Kwan MP, Wong MS, Huang J, Liu D. Identifying the space-time patterns of COVID-19 risk and their associations with different built environment features in Hong Kong. *Sci Total Environ*. 2019;2021(772):145379.
79. Hawkins D. Differential occupational risk for COVID-19 and other infection exposure according to race and ethnicity. *Am J Ind Med*. 2020;63:817–20.
80. Lan FY, Filler R, Mathew S, Buley J, Iliaki E, Bruno-Murtha LA, et al. Sociodemographic risk factors for coronavirus disease 2019 (COVID-19) infection among Massachusetts healthcare workers: A retrospective cohort study. *Infect Control Hosp Epidemiol*. 2021;42:1473–8.
81. Fauci AS, Lane HC, Redfield RR. Covid-19 — Navigating the Uncharted. *N Engl J Med*. 2020;382:1268–9.
82. Lamichhane DK, Shrestha S, Kim HC. District-level risk factors for COVID-19 incidence and mortality in Nepal. *Int J Environ Res Public Health*. 2022;19:2659.
83. Takahashi T, Ellingson MK, Wong P, Israelow B, Lucas C, Klein J, et al. Sex differences in immune responses that underlie COVID-19 disease outcomes. *Nature*. 2020;588:315–20.
84. Flook M, Jackson C, Vasileiou E, Simpson CR, Muckian MD, Agrawal U, et al. Informing the public health response to COVID-19: a systematic review of risk factors for disease, severity, and mortality. *BMC Infect Dis*. 2021;21:1–23.
85. Ayoub HH, Chemaitelly H, Mumtaz GR, Seedat S, Awad SF, Makhoul M, et al. Characterizing key attributes of COVID-19 transmission dynamics in China's original outbreak: model-based estimations. *Glob Epidemiol*. 2020;2:100042.
86. Hu S, Wang W, Wang Y, Litvinova M, Luo K, Ren L, et al. Infectivity, susceptibility, and risk factors associated with SARS-CoV-2 transmission under intensive contact tracing in Hunan, China. *Nat Commun*. 2021;12:1–11.
87. Yang X. Does city lockdown prevent the spread of COVID-19? New evidence from the synthetic control method. *Glob Health Res Policy*. 2021;6:20.
88. Sganzerla Martinez G, Hewins B, LeBlanc JJ, Ndishimye P, Toloue Ostadgavahi A, Kelvin DJ. Evaluating the effectiveness of lockdowns and restrictions during SARS-CoV-2 variant waves in the Canadian province of Nova Scotia. *Front Public Health*. 2023;11:1142602.
89. Erim DO, Oke GA, Adisa AO, Odukoya O, Ayo-Yusuf OA, Erim TN, et al. Associations of government-mandated closures and restrictions with aggregate mobility trends and SARS-CoV-2 infections in Nigeria. *JAMA Netw Open*. 2021;4:1–11.
90. Kazakos V, Taylor J, Luo Z. Impact of COVID-19 lockdown on NO2 and PM2.5 exposure inequalities in London, UK. *Environ Res*. 2021;198:111236.
91. Hohlfeld ASJ, Abdullahi L, Abou-Setta AM, Engel ME. International air travel-related control measures to contain the Covid-19 pandemic: a companion review to a Cochrane rapid review. *New Microbes New Infect*. 2022;49–50:101054.
92. Stutt ROJH, Retkute R, Bradley M, Gilligan CA, Colvin J. A modelling framework to assess the likely effectiveness of facemasks in combination with 'lock-down' in managing the covid-19 pandemic. *Proc R Soc A: Math, Phys Eng Sci*. 2020;476:20200376.
93. Anand U, Cabrerós C, Mal J, Ballesteros F, Sillanpää M, Tripathi V, et al. Novel coronavirus disease 2019 (COVID-19) pandemic: from transmission to control with an interdisciplinary vision. *Environ Res*. 2021;197:111126.
94. Nair AN, Anand P, George A, Mondal N. A review of strategies and their effectiveness in reducing indoor airborne transmission and improving indoor air quality. *Environ Res*. 2022;213:113579.
95. Bontempi E. Commercial exchanges instead of air pollution as possible origin of COVID-19 initial diffusion phase in Italy: more efforts are necessary to address interdisciplinary research. *Environ Res*. 2020;188:109775.
96. Bontempi E, Coccia M. International trade as critical parameter of COVID-19 spread that outclasses demographic, economic, environmental, and pollution factors. *Environ Res*. 2021;201:111514.
97. Bontempi E, Coccia M, Vergalli S, Zanoletti A. Can commercial trade represent the main indicator of the COVID-19 diffusion due to human-to-human interactions? A comparative analysis between Italy, France, and Spain. *Environ Res*. 2021;201:111529.
98. Dong YW, Liao ML, Meng XL, Somero GN. Structural flexibility and protein adaptation to temperature: molecular dynamics analysis of malate dehydrogenases of marine molluscs. *Proc Natl Acad Sci U S A*. 2018;115:1274–9.
99. Marti D, Torras J, Bertran O, Turon P, Alemán C. Temperature effect on the SARS-CoV-2: a molecular dynamics study of the spike homotrimeric glycoprotein. *Comput Struct Biotechnol J*. 2021;19:1848–62.
100. Marr LC, Tang JW, Van Mullekom J, Lakdawala SS. Mechanistic insights into the effect of humidity on airborne influenza virus survival, transmission and incidence. *J R Soc Interface*. 2019;16:20180298.
101. Schuit M, Biryukov J, Beck K, Yoltiz J, Bohannon J, Weaver W, et al. The stability of an isolate of the SARS-CoV-2 B.1.1.7 lineage in aerosols is similar to 3 earlier isolates. *J Infect Dis*. 2021;224:1641–8.
102. Shaman J, Kohn M. Absolute humidity modulates influenza survival, transmission, and seasonality. *Proc Natl Acad Sci U S A*. 2009;106:3243–8.
103. Shaman J, Pitzer VE, Viboud C, Grenfell BT, Lipsitch M. Absolute humidity and the seasonal onset of influenza in the continental United States. *PLoS Biol*. 2010;8:e1000316.
104. Kwon T, Gaudreault NN, Richt JA. Seasonal Stability of SARS-CoV-2 in Biological Fluids. *Pathogens*. 2021;10:540.
105. Kwon T, Gaudreault NN, Richt JA. Environmental stability of sars-cov-2 on different types of surfaces under indoor and seasonal climate conditions. *Pathogens*. 2021;10:1–8.
106. Pottage T, Onianwa O, Atkinson B, Spencer A, Bennett AM. Stability of SARS-CoV-2 variants of concern (Delta and Omicron) on surfaces at room temperature. *Virology*. 2023;583:27–8.
107. Guang Y, Hui L. Determining half-life of SARS-CoV-2 antigen in respiratory secretion. *Environ Sci Pollut Res*. 2023;30:69697–702.
108. Riddell S, Goldie S, Hill A, Eagles D, Drew TW. The effect of temperature on persistence of SARS-CoV-2 on common surfaces. *Virol J*. 2020;17:1–7.
109. Matson MJ, Yinda CK, Seifert SN, Bushmaker T, Fischer RJ, van Doremalen N, et al. Effect of environmental conditions on SARS-CoV-2 stability in human nasal mucus and sputum. *Emerg Infect Dis*. 2020;26:2276–8.
110. Kwon T, Gaudreault NN, Cool K, McDowell CD, Morozov I, Richt JA. Stability of SARS-CoV-2 in biological fluids of animals. *Viruses*. 2023;15:761.
111. Morris DH, Yinda KC, Gamble A, Rossine FW, Huang Q, Bushmaker T, et al. Mechanistic theory predicts the effects of temperature and humidity on inactivation of sars-cov-2 and other enveloped viruses. *Elife*. 2021;10:1–59.
112. Nocera AL, Mueller SK, Stephan JR, Hing L, Seifert P, Han X, et al. Exosome swarms eliminate airway pathogens and provide passive epithelial immunoprotection through nitric oxide. *J Allergy Clin Immunol*. 2019;143:1525–1535.e1.
113. Huang D, Taha MS, Nocera AL, Workman AD, Amiji MM, Bleier BS. Cold exposure impairs extracellular vesicle swarm-mediated nasal antiviral immunity. *J Allergy Clin Immunol*. 2023;151:509–525.e8.
114. Foxman EF, Storer JA, Fitzgerald ME, Wasik BR, Hou L, Zhao H, et al. Temperature-dependent innate defense against the common cold virus limits viral replication at warm temperature in mouse airway cells. *Proc Natl Acad Sci U S A*. 2015;112:827–32.
115. Kudo E, Song E, Yockey LJ, Rakib T, Wong PW, Homer RJ, et al. Low ambient humidity impairs barrier function and innate resistance against influenza infection. *Proc Natl Acad Sci U S A*. 2019;166:10905–10.
116. Walker CM, Ko G. Effect of ultraviolet germicidal irradiation on viral aerosols. *Environ Sci Technol*. 2007;41:5460–5.
117. Lorca-Oró C, Vila J, Pleguezuelos P, Vergara-Alert J, Rodon J, Majó N, et al. Rapid SARS-CoV-2 Inactivation in a simulated hospital room using a mobile and autonomous robot emitting ultraviolet-c light. *J Infect Dis*. 2022;225:587–92.

118. Schuit M, Ratnesar-Shumate S, Yoltz J, Williams G, Weaver W, Green B, et al. Airborne SARS-CoV-2 is rapidly inactivated by simulated sunlight. *J Infect Dis.* 2020;222:564–71.
119. Ratnesar-Shumate S, Williams G, Green B, Krause M, Holland B, Wood S, et al. Simulated sunlight rapidly inactivates SARS-CoV-2 on surfaces. *J Infect Dis.* 2020;222:214–22.
120. Sagripanti JL, Lytle CD. Estimated inactivation of coronaviruses by solar radiation with special reference to COVID-19. *Photochem Photobiol.* 2020;96:731–7.
121. O'Connor C, Courtney C, Murphy M. Shedding light on the myths of ultraviolet radiation in the COVID-19 pandemic. *Clin Exp Dermatol.* 2021;46:187–8.
122. Saraff V, Shaw N. Sunshine and vitamin D. *Arch Dis Child.* 2016;101:190–2.
123. Bogh MKB, Schmedes AV, Philipsen PA, Thieden E, Wulf HC. Vitamin D production depends on ultraviolet-B dose but not on dose rate: a randomized controlled trial. *Exp Dermatol.* 2011;20:14–8.
124. Bogh MKB. Vitamin D production after UVB: aspects of UV-related and personal factors. *Scand J Clin Lab Invest.* 2012;72(SUPPL. 243):24–31.
125. Pereira M, Dantas Damascena A, Galvão Azevedo LM, de Almeida OT, da Mota SJ. Vitamin D deficiency aggravates COVID-19: systematic review and meta-analysis. *Crit Rev Food Sci Nutr.* 2022;62:1308–16.
126. Cannell JJ, Vieth R, Umhau JC, Holick MF, Grant WB, Madronich S, et al. Epidemic influenza and vitamin D. *Epidemiol Infect.* 2006;134:1129–40.
127. Mohan M, Cherian JJ, Sharma A. Exploring links between vitamin D deficiency and covid-19. *PLoS Pathog.* 2020;16(9):e1008874.
128. Kumar D, Gupta P, Banerjee D. Letter: does vitamin D have a potential role against COVID-19? *Aliment Pharmacol Ther.* 2020;52:409–11.
129. Zheng SX, Yang JX, Hu X, Li M, Wang Q, Dancer RCA, et al. Vitamin D attenuates lung injury via stimulating epithelial repair, reducing epithelial cell apoptosis and inhibits TGF- $\beta$  induced epithelial to mesenchymal transition. *Biochem Pharmacol.* 2020;177:113955.
130. Mercola J, Grant WB, Wagner CL. Evidence regarding vitamin d and risk of covid-19 and its severity. *Nutrients.* 2020;12:1–24.
131. Lednický JA, Lauzard M, Fan ZH, Jutla A, Tilly TB, Gangwar M, et al. Viable SARS-CoV-2 in the air of a hospital room with COVID-19 patients. *Int J Infect Dis.* 2020;100:476–82.
132. Kutter JS, de Meulder D, Bestebroer TM, Lexmond P, Mulders A, Richard M, et al. SARS-CoV and SARS-CoV-2 are transmitted through the air between ferrets over more than one meter distance. *Nat Commun.* 2021;12:1653.
133. Wang CC, Prather KA, Sznitman J, Jimenez JL, Lakdawala SS, Tufekci Z, et al. Airborne transmission of respiratory viruses. *Science.* 2021;373:eabd9149.
134. Wei J, Li Y. Airborne spread of infectious agents in the indoor environment. *Am J Infect Control.* 2016;44:5102–8.
135. Chen W, Zhang N, Wei J, Yen HL, Li Y. Short-range airborne route dominates exposure of respiratory infection during close contact. *Build Environ.* 2020;176:106859.
136. Issakhov A, Omarova P, Abylkassymova A. Numerical simulation of social distancing of preventing airborne transmission in open space with lateral wind direction, taking into account temperature of human body and floor surface. *Environ Sci Pollut Res.* 2023;30:33206–28.
137. Spahn C, Hipp AM, Schubert B, Axt MR, Stratmann M, Schmölder C, et al. Airflow and Air velocity measurements while playing wind instruments, with respect to risk assessment of a SARS-CoV-2 infection. *Public Health.* 2021;18:5413.
138. Bhaganagar K, Bhimireddy S. Local atmospheric factors that enhance air-borne dispersion of coronavirus - high-fidelity numerical simulation of COVID19 case study in real-time. *Environ Res.* 2020;191:110170.
139. Motamedi H, Shirzadi M, Tominaga Y, Mirzaei PA. CFD modeling of airborne pathogen transmission of COVID-19 in confined spaces under different ventilation strategies. *Sustain Cities Soc.* 2022;76:103397.
140. Feng Y, Marchal T, Sperry T, Yi H. Influence of wind and relative humidity on the social distancing effectiveness to prevent COVID-19 airborne transmission: a numerical study. *J Aerosol Sci.* 2020;147:105585.
141. Chen W, Qian H, Zhang N, Liu F, Liu L, Li Y. Extended short-range airborne transmission of respiratory infections. *J Hazard Mater.* 2022;422:126837.
142. Bourdel T, Annesi-Maesano I, Alahmad B, Maesano CN, Bind MA. The impact of outdoor air pollution on covid-19: a review of evidence from in vitro, animal, and human studies. *Eur Respir Rev.* 2021;30:1–18.
143. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell.* 2020;181:271–280.e8.
144. Zhuang MW, Cheng Y, Zhang J, Jiang XM, Wang L, Deng J, et al. Increasing host cellular receptor—angiotensin-converting enzyme 2 expression by coronavirus may facilitate 2019-nCoV (or SARS-CoV-2) infection. *J Med Virol.* 2020;92:2693–701.
145. Zhou L, Xu Z, Castiglione GM, Soiberman US, Eberhart CG, Duh EJ. ACE2 and TMPRSS2 are expressed on the human ocular surface, suggesting susceptibility to SARS-CoV-2 infection. *Ocular Surface.* 2020;18:537–44.
146. Li HH, Liu CC, Hsu TW, Lin JH, Hsu JW, Li AFY, et al. Upregulation of ACE2 and TMPRSS2 by particulate matter and idiopathic pulmonary fibrosis: a potential role in severe COVID-19. *Part Fibre Toxicol.* 2021;18:1–13.
147. Sagawa T, Tsujikawa T, Honda A, Miyasaka N, Tanaka M, Kida T, et al. Exposure to particulate matter upregulates ACE2 and TMPRSS2 expression in the murine lung. *Environ Res.* 2020;2021(1195):110722.
148. Chuang HC, Chen YY, Hsiao TC, Chou HC, Kuo HP, Feng PH, et al. Alteration in angiotensin-converting enzyme 2 by pm1 during the development of emphysema in rats. *ERJ Open Res.* 2020;6:1–8.
149. Wang H, Song L, Ju W, Wang X, Dong L, Zhang Y, et al. The acute airway inflammation induced by PM 2.5 exposure and the treatment of essential oils in Balb/c mice. *Sci Rep.* 2017;7:44256.
150. Deng L, Ma M, Li S, Zhou L, Ye S, Wang J, et al. Protective effect and mechanism of baicalin on lung inflammatory injury in BALB/C mice induced by PM2.5. *Ecotoxicol Environ Saf.* 2022;248:114329.
151. Du X, Jiang S, Zeng X, Zhang J, Pan K, Song L, et al. Fine particulate matter-induced cardiovascular injury is associated with NLRP3 inflammasome activation in Apo E<sup>-/-</sup> mice. *Ecotoxicol Environ Saf.* 2018;2019(174):92–9.
152. Kampfrath T, Maiseyeu A, Ying Z, Shah Z, Deilüls JA, Xu X, et al. Chronic fine particulate matter exposure induces systemic vascular dysfunction via NADPH oxidase and TLR4 pathways. *Circ Res.* 2011;108:716–26.
153. Riva DR, Magalhães CB, Lopes AA, Lanças T, Mauad T, Malm O, et al. Low dose of fine particulate matter (PM2.5) can induce acute oxidative stress, inflammation and pulmonary impairment in healthy mice. *Inhal Toxicol.* 2011;23:257–67.
154. Wang S, Zhou Q, Tian Y, Hu X. The lung microbiota affects pulmonary inflammation and oxidative stress induced by PM2.5 exposure. *Environ Sci Technol.* 2022;56:12368–79.
155. Martin PJ, Billet S, Landkocz Y, Fougère B. Inflammation at the crossroads: the combined effects of COVID-19, ageing, and air pollution. *J Frailty Aging.* 2021;10:281–5.
156. Wang B, Chen H, Yik X, Chan L, Oliver BG. Is there an association between the level of ambient air pollution and COVID-19? *J Physiol Lung Cell Mol Physiol.* 2020;319:416–21.
157. Liu Q, Weng J, Li C, Feng Y, Xie M, Wang X, et al. Attenuation of PM2.5-induced alveolar epithelial cells and lung injury through regulation of mitochondrial fission and fusion. *Part Fibre Toxicol.* 2023;20:28.
158. To EE, Vlahos R, Luong R, Halls ML, Reading PC, King PT, et al. Endosomal NOX2 oxidase exacerbates virus pathogenicity and is a target for antiviral therapy. *Nat Commun.* 2017;8:69.
159. Sciomer S, Moscucci F, Magri D, Badagliacca R, Piccirillo G, Agostoni P. SARS-CoV-2 spread in Northern Italy: what about the pollution role? *Environ Monit Assess.* 2020;192:2–4.
160. Conticini E, Frediani B, Caro D. Can atmospheric pollution be considered a co-factor in extremely high level of SARS-CoV-2 lethality in Northern Italy? *Environ Pollut.* 2020;261:114465.
161. Miller MR. Oxidative stress and the cardiovascular effects of air pollution. *Free Radical Biol Med.* 2020;151:69–87.
162. Bromberg PA. Mechanisms of the acute effects of inhaled ozone in humans. *Biochim Biophys Acta Gen Subj.* 2016;1860:2771–81.
163. Wiegman CH, Li F, Ryffel B, Togbe D, Chung KF. Oxidative stress in ozone-induced chronic lung inflammation and emphysema: a facet of chronic obstructive pulmonary disease. *Front Immunol.* 2020;11:1957.
164. Manzer R, Dinarello CA, McConville G, Mason RJ. Ozone exposure of macrophages induces an alveolar epithelial chemokine response through IL-1 $\alpha$ . *Am J Respir Cell Mol Biol.* 2008;38:318–23.



165. Chen QZ, Zhou YB, Zhou LF, Fu ZD, Wu YS, Chen Y, et al. TRPC6 modulates adhesion of neutrophils to airway epithelial cells via NF- $\kappa$ B activation and ICAM-1 expression with ozone exposure. *Exp Cell Res*. 2019;377:56–66.
166. Wiegman CH, Li F, Clarke CJ, Jazrawi E, Kirkham P, Barnes PJ, et al. A comprehensive analysis of oxidative stress in: the ozone-induced lung inflammation mouse: model. *Clin Sci*. 2014;126:425–40.
167. Bauer AK, Rondini EA, Hummel KA, Degraff LM, Walker C, Jedlicka AE, et al. Identification of candidate genes downstream of TLR4 signaling after ozone exposure in mice: A role for heat-shock protein 70. *Environ Health Perspect*. 2011;119:1091–7.
168. Mumby S, Chung KF, Adcock IM. Transcriptional effects of ozone and impact on airway inflammation. *Front Immunol*. 2019;10:1610.
169. Paludan SR, Reinert LS, Hornung V. DNA-stimulated cell death: implications for host defence, inflammatory diseases and cancer. *Nat Rev Immunol*. 2019;19:141–53.
170. Zhang JH, Yang X, Chen YP, Zhang JF, Li CQ. Nrf2 activator RTA-408 Protects against ozone-induced acute asthma exacerbation by suppressing ROS and  $\gamma\delta$ T17 cells. *Inflammation*. 2019;42:1843–56.
171. Fry RC, Rager JE, Zhou H, Zou B, Brickey JW, Ting J, et al. Individuals with increased inflammatory response to ozone demonstrate muted signaling of immune cell trafficking pathways. *Respir Res*. 2012;13:89.
172. Bao A, Yang H, Ji J, Chen Y, Bao W, Li F, et al. Involvements of p38 MAPK and oxidative stress in the ozone-induced enhancement of AHR and pulmonary inflammation in an allergic asthma model. *Respir Res*. 2017;18:216.
173. Mazur-Panasiuk N, Botwina P, Kutaj A, Woszczyna D, Pyrc K. Ozone treatment is insufficient to inactivate sars-cov-2 surrogate under field conditions. *Antioxidants*. 2021;10:1480.
174. Tizaoui C. Ozone: A Potential Oxidant for COVID-19 Virus (SARS-CoV-2). *Ozone Sci Eng*. 2020;42:378–85.
175. Tizaoui C, Stanton R, Statkute E, Rubina A, Lester-Card E, Lewis A, et al. Ozone for SARS-CoV-2 inactivation on surfaces and in liquid cell culture media. *J Hazard Mater*. 2022;428:128251.
176. Elvis AM, Ekta JS. Ozone therapy: a clinical review. *J Nat Sci Biol Med*. 2011;2:66–70.
177. Cattel F, Giordano S, Bertiond C, Lupia T, Corcione S, Scaldaferrri M, et al. Ozone therapy in COVID-19: a narrative review. *Virus Res*. 2021;291:198207.
178. Sagai M, Bocci V. Mechanisms of action involved in ozone therapy: is healing induced via a mild oxidative stress? *Med Gas Res*. 2011;1:29.
179. Amor S, Fernández Blanco L, Baker D. Innate immunity during SARS-CoV-2: evasion strategies and activation trigger hypoxia and vascular damage. *Clin Exp Immunol*. 2020;202:193–209.
180. Bocci V, Borrelli E, Travagli V, Zanardi I. The ozone paradox: Ozone is a strong oxidant as well as a medical drug. *Med Res Rev*. 2009;29:646–82.
181. Serra MEG, Baeza-Noci J, Abdala CVM, Luvisotto MM, Bertol CD, Anzolin AP. Clinical effectiveness of medical ozone therapy in COVID-19: the evidence and gaps map. *Med Gas Res*. 2023;13:172–80.
182. Villeneuve PJ, Goldberg MS. Methodological considerations for epidemiological studies of air pollution and the sars and COVID-19 coronavirus outbreaks. *Environ Health Perspect*. 2020;128:95001.
183. Benmarhnia T. Linkages between air pollution and the health burden from covid-19: methodological challenges and opportunities. *Am J Epidemiol*. 2020;189:1238–43.
184. Shakil MH, Munim ZH, Tasnia M, Sarowar S. COVID-19 and the environment: a critical review and research agenda. *Sci Total Environ*. 2020;745:141022.
185. D'Amico F, Marmiere M, Righetti B, Scquizzato T, Zangrillo A, Puglisi R, et al. COVID-19 seasonality in temperate countries. *Environ Res*. 2021;2022(206):112614.
186. Jiang C, Wang X, Li X, Inlora J, Wang T, Liu Q, et al. Dynamic human environmental exposome revealed by longitudinal personal monitoring. *Cell*. 2018;175:277–291.e31.
187. Beltran RM, Holloway IW, Hong C, Miyashita A, Cordero L, Wu E, et al. Social determinants of disease: HIV and COVID-19 experiences. *Curr HIV/AIDS Rep*. 2022;19:101–12.
188. Pan A, Liu L, Wang C, Guo H, Hao X, Wang Q, et al. Association of public health interventions with the epidemiology of the COVID-19 outbreak in Wuhan, China. *JAMA - J Am Med Asso*. 2020;323:1915–23.
189. Telenti A, Arvin A, Corey L, Corti D, Diamond MS, García-Sastre A, et al. After the pandemic: perspectives on the future trajectory of COVID-19. *Nature*. 2021;596:495–504.
190. Kurt OK, Zhang J, Pinkerton KE. Pulmonary health effects of air pollution. *Curr Opin Pulm Med*. 2016;22:138–43.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.